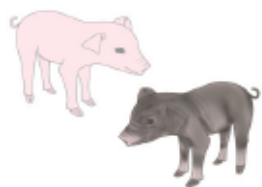


Quelques exemples avec des données multi-omiques et multi-tissus en néonatalité porcine pour illustrer les différentes fonctionnalités d'Asterics.

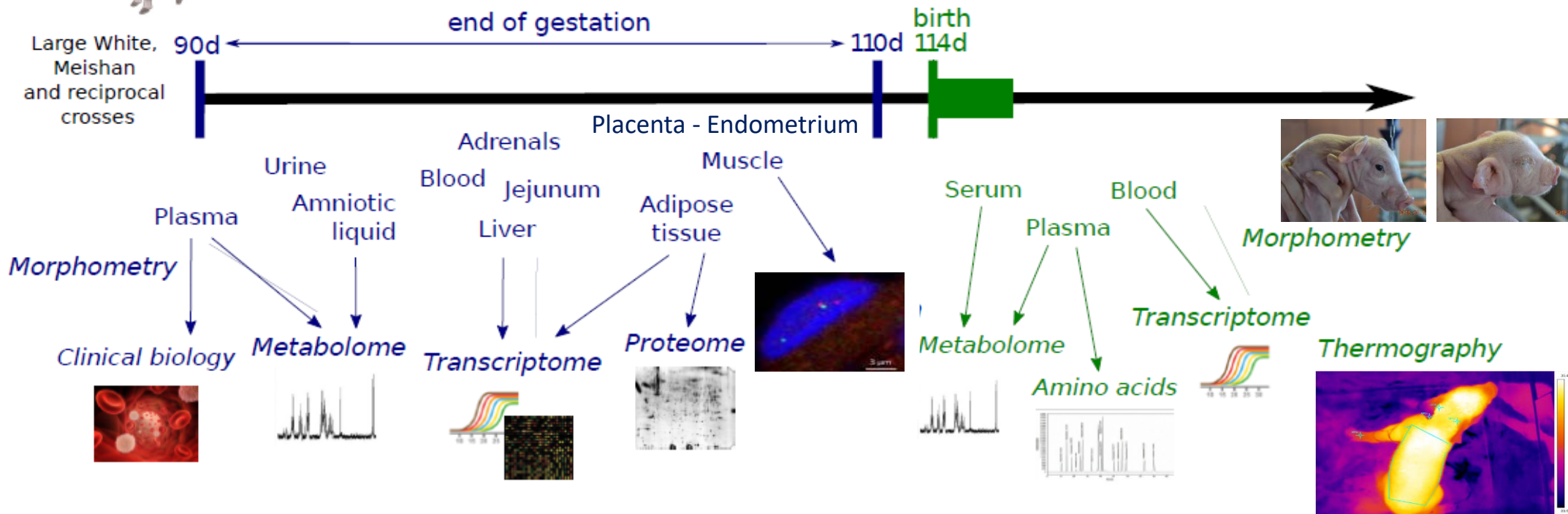
Élise Maigné, Céline Noirot, Jérôme Mariette, Yaa Adu Kesewaah, Sébastien Déjean, Camille Guilmineau, Julien Henry, Arielle Krebs, **Laurence Liaubet**, Fanny Mathevet, Hyphen-Stat, Christine Gaspin, Nathalie Vialaneix



Context: neonatal maturity and birth survival



3 stages: 90 or 110 days of gestation, birth
 4 genotypes: LW, MS, LWMS, MSLW
 N: 50 ... 600... 900



Voillet et al, 2014, 2018; Yao et al.2017 ; Gondret et al, 2018;
 Marti-Marimon et al, 2018; Lefort et al, 2020,2021 ; Schmitt et al, 2021



asterics

A tool for the exploration
and integration of **omics data**

ASTERICS is an online tool designed to help you to perform
your statistical and integrative analyses in an interactive
and easy-to-use way.




[+ Create new workspace](#) [Load workspace](#)

Load TCGA demo data into your workspace

I confirm that I have read and that I agree to *Asterics' Privacy Policy*.

[+ Create new workspace](#)

 Your unique identifier will allow you to access your work and data during 30 days after the last activity on ASTERICS.

<https://asterics.miat.inrae.fr/>

Project coordinators



Funder



Partners



Dear user,

Your workspace has been successfully created at

<https://asterics.miat.inrae.fr/workspace/ba20fb6d-4196-45c0-999f-7ff6d522b109>.

Thank you for using ASTERICS,
The ASTERICS Team

Dear user,

your workspace <https://asterics.miat.inrae.fr/78f3057f-ed70-42c8-ae5c-8d8b7be83484> will be deleted in 3 days.

thanks for using Asterics,
The Asterics Team,

 Datasets 0 Variables 0 Analyses 0 Remaining days 30

Copy url

 Workflow All datasets Remove Add

| <input type="checkbox"/> | Name | Type | # Rows | # Columns | Log | Normalized |
|--------------------------|------|------|--------|-----------|-----|------------|
|--------------------------|------|------|--------|-----------|-----|------------|



Nothing's here...

 All analyses Remove Export report

| <input type="checkbox"/> | Name | Function |
|--------------------------|------|----------|
|--------------------------|------|----------|



Nothing's here...

1 Introduction**2** Workspace

2.1 Create / load a workspace

2.2 Screen "My workspace"

2.3 Import your data

2.4 Retrieve an analysis

2.5 Export the data

2.6 Export report

3 What you can do with your data

3.1 Edit

3.2 Explore

3.3 Integrate

I Edit**4** Edit a dataset**5** Transformation

5.1 Standard transformation

5.2 Quantile normalization

5.3 RNA-seq (and other count) d...

5.4 Compositional data

5.5 Metagenomic data

5.6 Correcting batch effect with ...

6 Missing values

6.1 Explore missing values

6.2 Remove missing values

6.3 Impute missing values

II Explore**7** Explore variables

7.1 Univariate

7.2 Bivariate

ASTERICS: User documentation

Maigné Élise, Noirot Céline, Mariette Jérôme, Adu Kesewaah Yaa, Déjean Sébastien,

Guilmineau Camille, Henry Julien, Krebs Arielle, Liaubet Laurence, Mathevet Fanny,

Hyphen-Stat, Gaspin Christine, Vialaneix Nathalie

2022-08-17

Section 1 Introduction

This document is the user guide for the web tool [ASTERICS](#). *It is still a work-in-progress and mostly incomplete. Come back later for a better version. Thank you for your understanding.*

Link to the application: <http://asterics.miat.inrae.fr/>

Support for [ASTERICS](#) can be obtained at asterics-tlse@inrae.fr.

Bugs can be reported [here](#) and suggestions can be made [here](#).



asterics

A tool for the exploration and integration of omics data

- 13.2 Run PLS-DA
- 13.3 Explore individuals
- 13.4 Explore variables
- 13.5 Extract new data
- 14 Multiple Factor Analysis (MFA)
 - 14.1 Preprocessing
 - 14.2 Run MFA
 - 14.3 Explore individuals
 - 14.4 Explore variables
 - 14.5 Explore groups
 - 14.6 Extract new data
- 15 Differential Analysis
 - 15.1 Preprocessing
 - 15.2 Multiple tests
 - 15.3 Posthoc tests
 - 15.4 Extract dataset
- IV Case studies
 - 16 Case studies
 - 16.1 Breast cancer
 - 16.1.1 Setup
 - 16.1.2 Edition
 - 16.1.3 Principal Component A...
 - 16.1.4 Hierarchical clustering
 - 16.1.5 Data import
 - 16.1.6 Normalization
 - 16.1.7 Multiple Factor Analy...
 - 16.1.8 Partial Least Squares -...
 - 16.1.9 Interpretation
- References

Section 16 Case studies

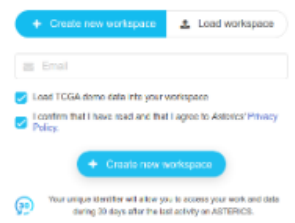
16.1 Breast cancer

This case study focuses on breast cancer, trying to link omics data to cancer subtypes. There are four breast cancer subtypes that somewhat represent sub-diseases, with different biological mechanisms. Their identification is particularly important when designing / selecting treatments. For this case study, we have mRNA and miRNA data on 988 individuals, as well as the associated subtypes. mRNA data is already normalized and is loaded in ASTERICS by default, whereas miRNA comes in the form of raw counts.

The data is available online ([Vialaneix 2021](#)).

16.1.1 Setup

Let us open an empty session and load the TCGA demo data.



Since we will only use the `mrna` dataset, let us delete the other two.

All datasets Remove Add

| <input type="checkbox"/> | Name | Type | # Rows | # Columns | Log | Normalized | |
|-------------------------------------|----------|----------|--------|-----------|-----|------------|--|
| <input type="checkbox"/> | mrna | ma-count | 988 | 2000 | yes | yes | |
| <input checked="" type="checkbox"/> | protein | generic | 379 | 142 | no | no | |
| <input checked="" type="checkbox"/> | clinical | metadata | 989 | 217 | no | no | |

Here is `mrna` in the workspace:

```
mrna
ma-count
#rows: 988 #col.: 2000
```

1 File upload

2 Dataset structure

1. Select a file to upload

File

Select a file

Dataset name

Dataset name

The dataset name field is required

Dataset nature

* Dataset nature

- generic
- sequencing data (RNAseq, miRNA, ...) counts
- microarray transcriptomics**
- metagenomics count data
- metagenomics composition (compositional data)
- metabolite/protein/lipid composition (compositional data)
- phenotypes
- metadata (or design)
- other

If you are not sure!

Before importation

Have data been log-transformed?

Have data been normalized?

Q Raw file

Select a file to get a preview

1. Select a file to upload



File

 TranscriptomeMuscle50.csv

Dataset name

 TranscriptomeMuscle50 

Dataset nature

 microarray transcriptomics 

If you don't know which nature to choose, use "generic".

Before importation

Have data been log-transformed?

Have data been normalized?

Q Raw file

```
;F405;F409;F411;F449;F455;F458;F469;F476;F481;F485;F498;F500;F524;F525;F53
A1CF_52154;5.6803695954516;5.45610320624039;5.74484272573927;5.92019267767
A2M_40313;5.50693239097091;5.76073759718855;5.5227941006065;5.663677602194
A2M_10645;5.41017364325691;5.79477650004684;6.04237760287688;5.68692117450
A2ML1_14639;5.4491082475774;5.36385961056552;5.17100665288747;5.6208513292
A4GALT_23662;8.36976136324847;7.80328185134206;8.55421636844331;7.22995624
A4GALT_9159;8.82297280814355;8.92285084244449;9.17229076620426;8.834796301
AAAS_61230;5.66491543991508;5.70809268946484;5.85667371223367;6.9757878359
AAAS_2894;5.89283524674721;5.6564212452558;5.66624190687793;6.090788421745
AAAS_18735;7.74349641324072;7.70023645818195;7.43741001836278;8.7585108190
AAAS_24237;6.41516988709249;6.40535808528583;6.82967565563798;7.0901431036
```



2. Set the dataset structure

Column names are included in the first row

File has row labels

Index of the column with row labels

1

Comment character

#

File has comment lines

Switch rows and columns

For ASTERICS, individuals must be in rows.

Separator

, (comma) ; (semicolon) \t (tab) Space Custom:

Quote mark for text

" (double quote)

Decimal mark

. (dot) , (comma)

Encoding

Auto UTF-8 latin1

Missing values are encoded with these strings:

NA Add na.strings

Be patient: large files might take a long time to be uploaded and processed!



Import

Q Raw file

```
;F476;F481;F560;F578;F688;F455;F485;F498;F738;F899;F900;F469;F557;F559;F67
A1CF_52154;5.82599980707194;5.91662162096354;5.62770189995087;5.9748791175
A2M_40313;5.20290966528126;5.71662080795284;5.73529430299502;6.27994060037
A2M_10645;6.05043272137919;5.78608436449946;5.92397285829613;6.31840463378
A2ML1_14639;5.09454305674891;4.89392730422894;5.06610396900029;5.109927232
A4GALT_23662;7.44499238746051;7.88769803161222;7.75157714142366;7.85478806
A4GALT_9159;8.80734586555401;8.74392533000205;8.55935004010872;8.291677946
AAAS_61230;5.82599980707194;5.60366933006277;6.19088825000998;6.6908486193
AAAS_2894;5.59207508123505;5.29881778254516;5.87420980899597;5.93550088274
AAAS_18735;7.9656631095485;7.65139989082614;7.78415314320795;8.09455028881
AAAS_24237;6.38767805826733;6.53943474153246;6.73107621612731;6.5142754375
```

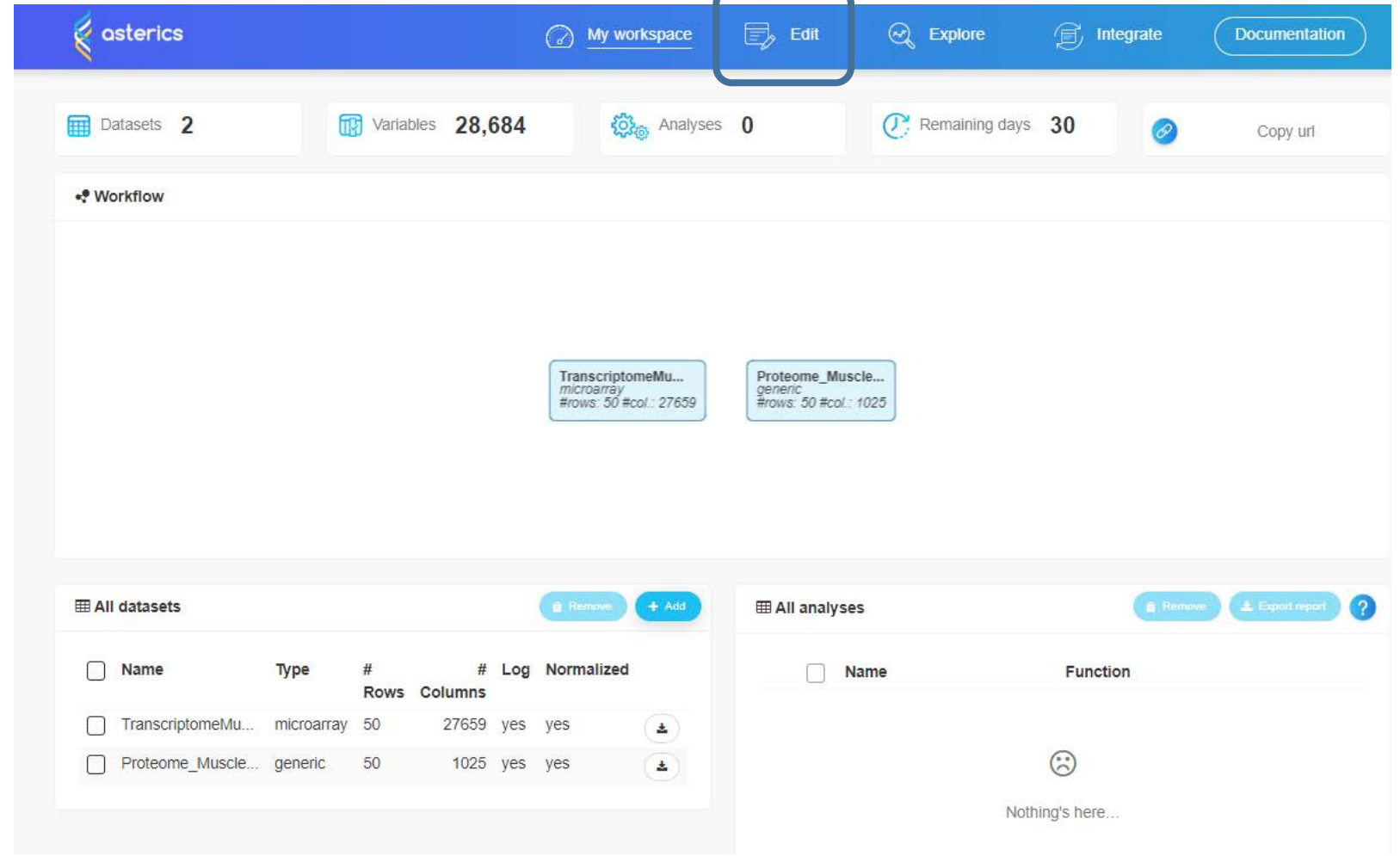
Switch rows and columns !

Q Dataset preview

| | A1CF_52154 numeric | A2M_40313 numeric | A2M_10645 numeric | A2ML1_14639 numeric | A4GALT_23662 numeric | A4GALT_9159 numeric |
|------|-----------------------|----------------------|----------------------|------------------------|-------------------------|------------------------|
| F476 | 5.826 | 5.2029 | 6.0504 | 5.0945 | 7.445 | |
| F481 | 5.9166 | 5.7166 | 5.7861 | 4.8939 | 7.8877 | |
| F560 | 5.6277 | 5.7353 | 5.924 | 5.0661 | 7.7516 | |
| F578 | 5.9749 | 6.2799 | 6.3184 | 5.1099 | 7.8548 | |
| F688 | 5.7889 | 6.434 | 6.5869 | 4.7516 | 8.0703 | |
| F455 | 5.5397 | 5.5826 | 5.8344 | 5.648 | 7.4885 | |
| F485 | 5.8868 | 6.1856 | 6.3453 | 5.661 | 7.7702 | |
| F498 | 5.6354 | 5.81 | 5.7552 | 5.8236 | 7.6827 | |
| F738 | 5.6221 | 5.1073 | 5.5711 | 5.6786 | 7.1388 | |
| F899 | 5.5986 | 5.5457 | 5.8591 | 5.0093 | 7.3409 | |
| F900 | 5.781 | 5.9127 | 6.1895 | 5.8663 | 7.7117 | |
| F469 | 6.1845 | 5.7959 | 5.7232 | 5.5926 | 8.0633 | |
| F557 | 5.8742 | 5.8665 | 6.051 | 5.5620 | 7.0101 | |

1 unique identifier per variable, so arrange the name to have the most relevant information !

Edit

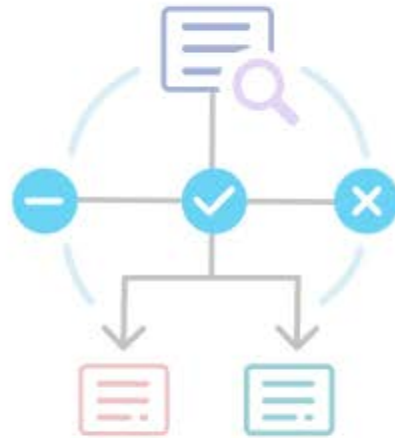


The screenshot shows the asterics interface with the following components:

- Navigation Bar:** Contains the asterics logo, 'My workspace', 'Edit' (highlighted with a blue box and arrow), 'Explore', 'Integrate', and 'Documentation' buttons.
- Summary Cards:** Datasets: 2, Variables: 28,684, Analyses: 0, Remaining days: 30, and a 'Copy url' button.
- Workflow Section:** Displays two dataset cards: 'TranscriptomeMu... microarray #rows: 50 #col.: 27659' and 'Proteome_Muscle... generic #rows: 50 #col.: 1025'.
- All datasets Table:**

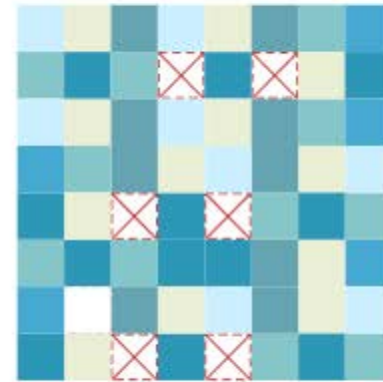
| <input type="checkbox"/> | Name | Type | # Rows | # Columns | Log | Normalized | |
|--------------------------|--------------------|------------|--------|-----------|-----|------------|--|
| <input type="checkbox"/> | TranscriptomeMu... | microarray | 50 | 27659 | yes | yes | |
| <input type="checkbox"/> | Proteome_Muscle... | generic | 50 | 1025 | yes | yes | |
- All analyses Section:** Shows a table with columns 'Name' and 'Function', currently empty with a sad face icon and the text 'Nothing's here...'.

Let's edit data!



Dataset edition

Provides several ways to modify a dataset.



Missing values

Explore, remove or impute missing values on a dataset.



Normalize dataset

Correct technical biases and prepare your data for further analyses.

Dataset edition

1. Select a dataset and an action ?

Dataset

Infos_50 ▼

Actions

- Transpose
- Change dataset nature
- Change variable (column) types
- Set individual (row) names
- Subset individuals (rows)
- Subset variables (columns)
- Rename categories
- Reorder categories

History

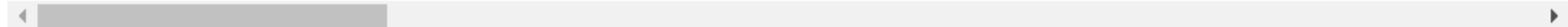
No history to display

Select an action.

Dataset "Infos_50"

| # rows | # col. | # missing | % missing | # numeric | # cat. | # logic | # others |
|--------|--------|-----------|-----------|-----------|--------|---------|----------|
| 50 | 32 | 0 | 0 | 27 | 5 | 0 | 0 |

| | Sex categorical 0.00% | N_Mother numeric 0.00% | Age categorical 0.00% | TG_Mother categorical 0.00% | TG_Father categorical 0.00% | TG_F categorical 0.00% | Weight numeric 0.00% |
|------|-----------------------------|------------------------------|-----------------------------|-----------------------------------|-----------------------------------|------------------------------|----------------------------|
| F449 | M | | 6 90d | LW | LW | LWLW | 733 |
| F458 | M | | 6 90d | LW | LW | LWLW | 782 |
| F500 | M | | 8 90d | LW | LW | LWLW | 597 |
| F736 | M | | 23 90d | LW | LW | LWLW | 597 |
| F744 | M | | 23 90d | LW | LW | LWLW | 784 |
| F894 | M | | 33 90d | LW | LW | LWLW | 813 |
| F895 | M | | 33 90d | LW | LW | LWLW | 389 |
| F455 | M | | 6 90d | LW | MS | MSLW | 717 |
| F485 | M | | 8 90d | LW | MS | MSLW | 754 |
| F498 | M | | 8 90d | LW | MS | MSLW | 704 |
| F738 | M | | 23 90d | LW | MS | MSLW | 522 |
| F899 | M | | 33 90d | LW | MS | MSLW | 599 |



Explore variables in a dataset

1 variable 2 variables up to 5 variables

all variables in a dataset

1. Select the dataset to explore

Select a dataset

Infos_50_edited

Scale numeric variables in the plot

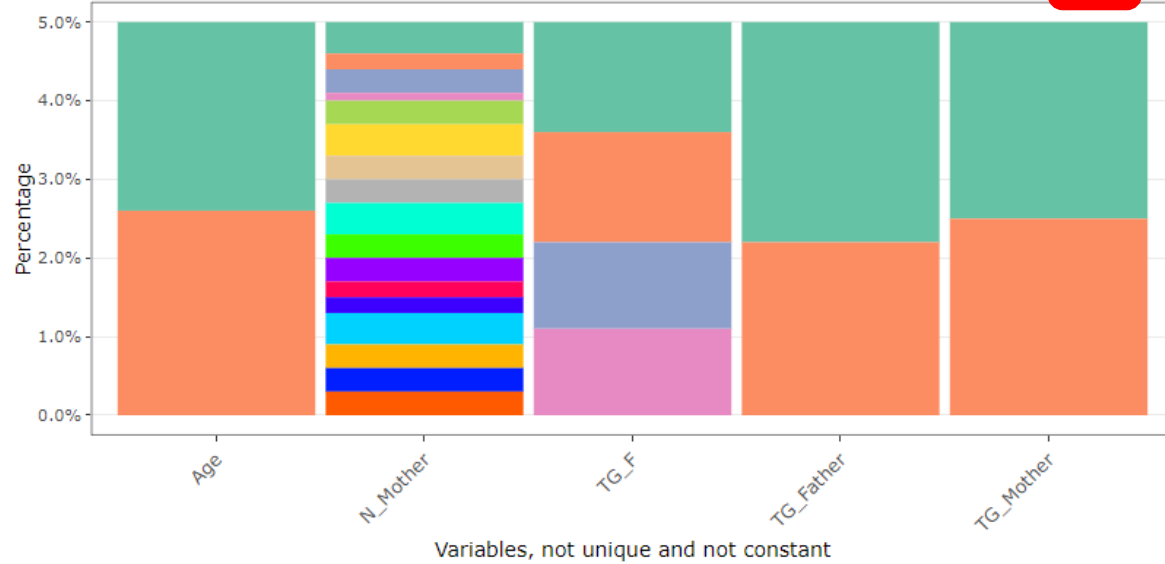
Explore

2. Plots and numerical summaries

One irrelevant categorical variable (Sex) has been removed from the dataset because it had a unique value or only different values.

Numeric summary Normality test Categorical summary Plots

Categorical variables of Infos_50_edited



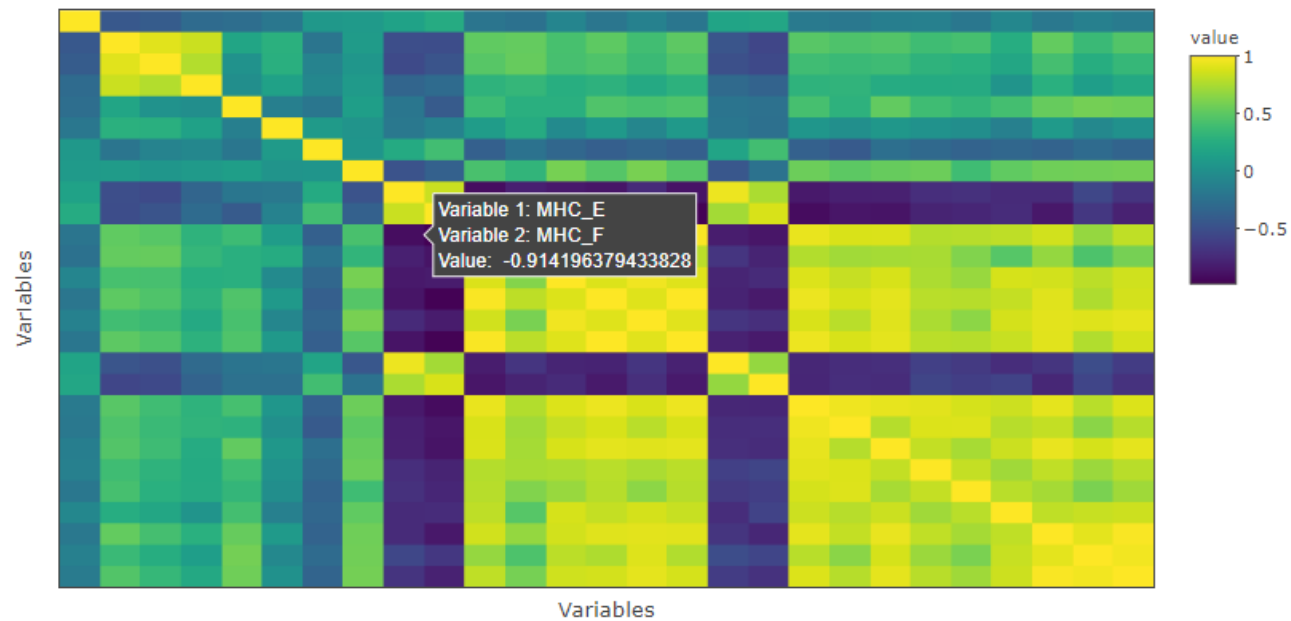
1 variable 2 variables up to 5 variables **all variables in a dataset**

2. Plots and numerical summaries

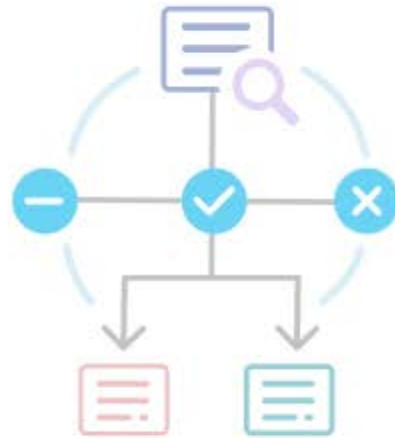
One irrelevant categorical variable (Sex) has been removed from the dataset because it had a unique value or only different values.

 Numeric summary  Normality test  Categorical summary  Plots

Correlation plot of Intos_50

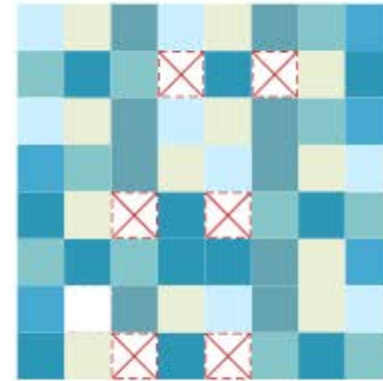


Let's edit data!



Dataset edition

Provides several ways to modify a dataset.



Missing values

Explore, remove or impute missing values on a dataset.



Normalize dataset

Correct technical biases and prepare your data for further analyses.

Which decisions should I take to handle missing values?

qPCR data...



| # rows | # col. | # missing | % missing | # numeric | # cat. | # logic | # others |
|--------|--------|-----------|-----------|-----------|--------|---------|----------|
| 257 | 45 | 2760 | 23.87 | 45 | 0 | 0 | 0 |

| | ATP2B1 numeric 1.17% | BCAS2 numeric 38.52% | BIRC5_F.R numeric 56.03% | CD14 numeric 39.30% | CD3E numeric 0.39% | CIR1 numeric 34.63% | CSNK2A1 numeric 79.38% | |
|----|----------------------------|----------------------------|--------------------------------|---------------------------|--------------------------|---------------------------|------------------------------|----|
| P1 | 0.0559 | 0.0623 | NA | 0.1167 | 0.0191 | 0.0132 | NA | NA |
| P2 | 0.0521 | 0.1207 | NA | 0.042 | 0.0828 | 0.0399 | NA | NA |
| P3 | 0.153 | 0.0737 | NA | 0.0761 | 0.0773 | NA | 0.0826 | NA |
| P4 | 0.0644 | 0.1384 | NA | 0.2347 | 0.0495 | 0.0555 | NA | NA |
| P5 | 0.2821 | 0.246 | NA | 0.3472 | 0.2379 | 0.0441 | NA | NA |
| P6 | 0.1404 | 0.1411 | 0.1522 | 0.0928 | 0.0943 | NA | NA | NA |
| P7 | 0.5945 | NA | 0.1526 | NA | 0.2019 | 0.3797 | NA | NA |
| P8 | 0.1211 | 0.1025 | 0.088 | 0.1648 | 0.141 | NA | NA | NA |

Missing values

[Explore missing values](#)[Remove missing values](#)[Impute missing values](#)

1. Select the dataset to analyze

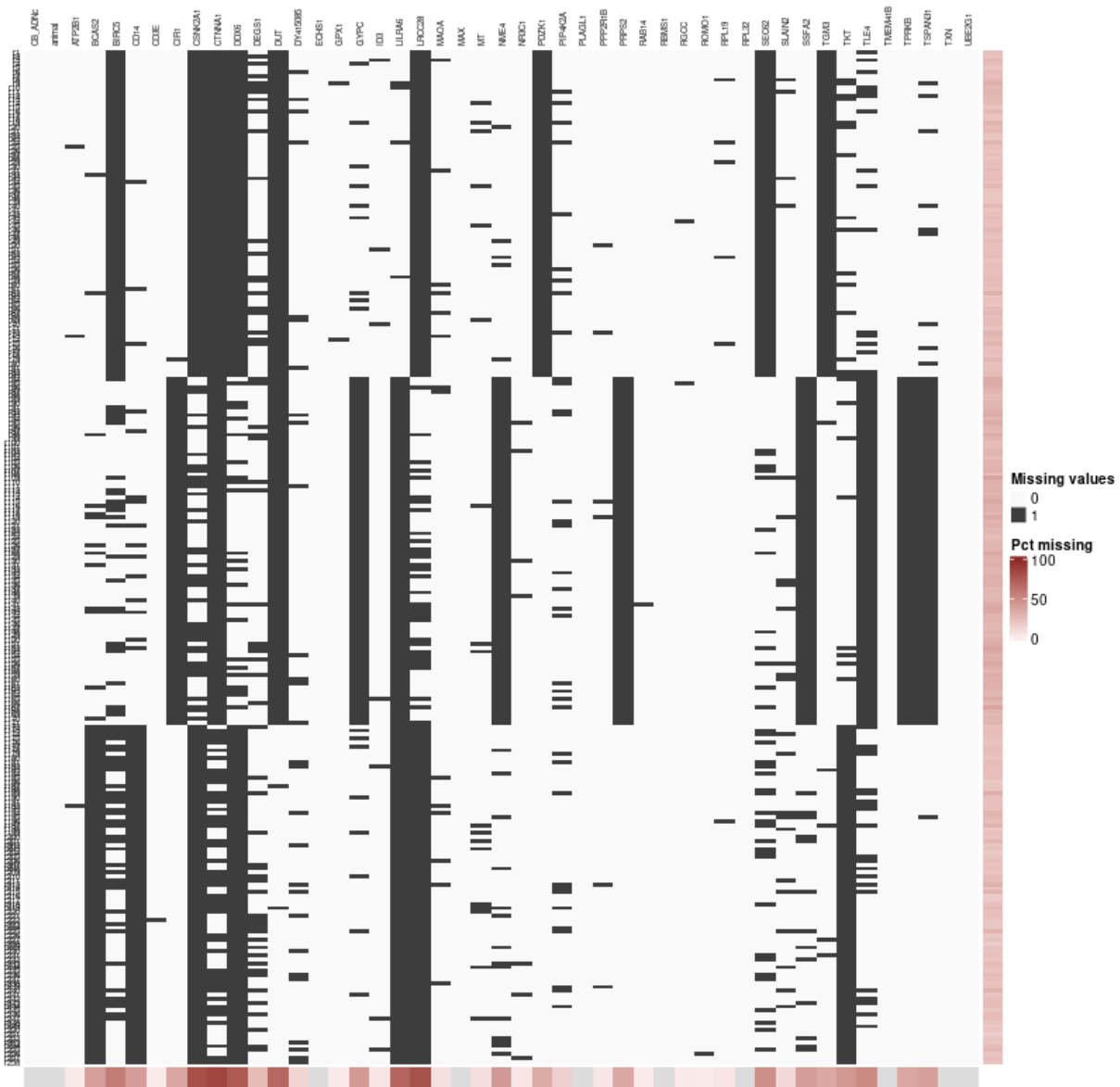
Select a dataset

 PICLET_qPCR1P218 ▾

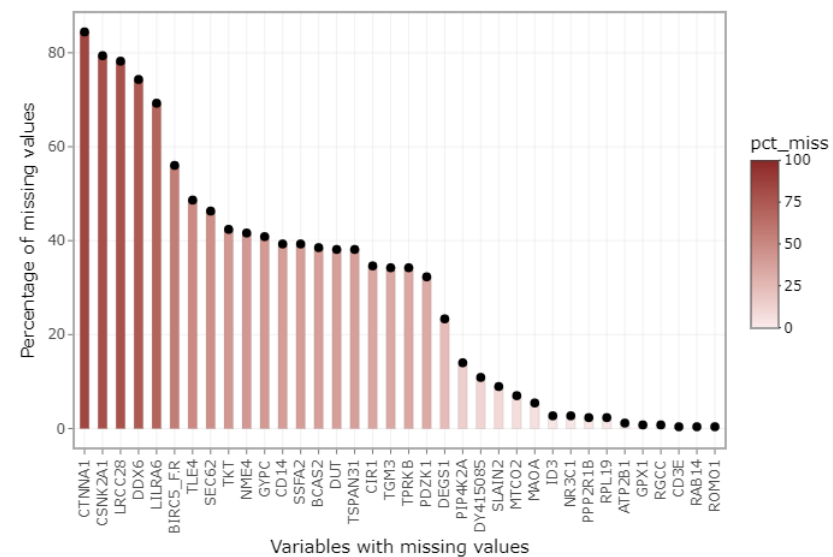
2. Plots and numerical summaries

[Heatmaps](#)[Plots](#)[Summary](#)

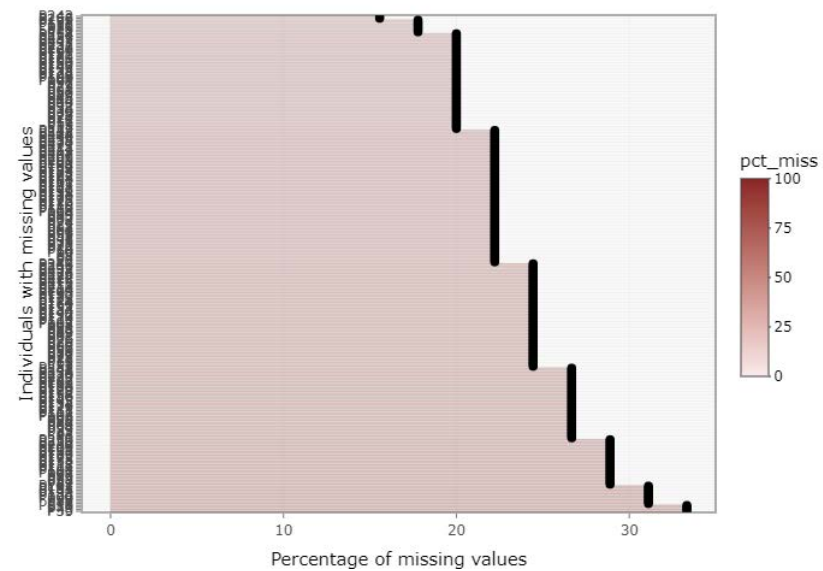
qPCR-1: Heatmap of missing values



PICLET_qPCR1P218: Percentage of missing values by variable



PICLET_qPCR1P218: Percentage of missing values by individual



Missing values

[Explore missing values](#)
[Remove missing values](#)
[Impute missing values](#)

1. Select the dataset to analyze ?

Select a dataset

Direction

Maximum % of acceptable missingness in individuals/variables

Run

2. Plots and numerical summaries

Purpose of Missing values:

Handle missing values in your datasets

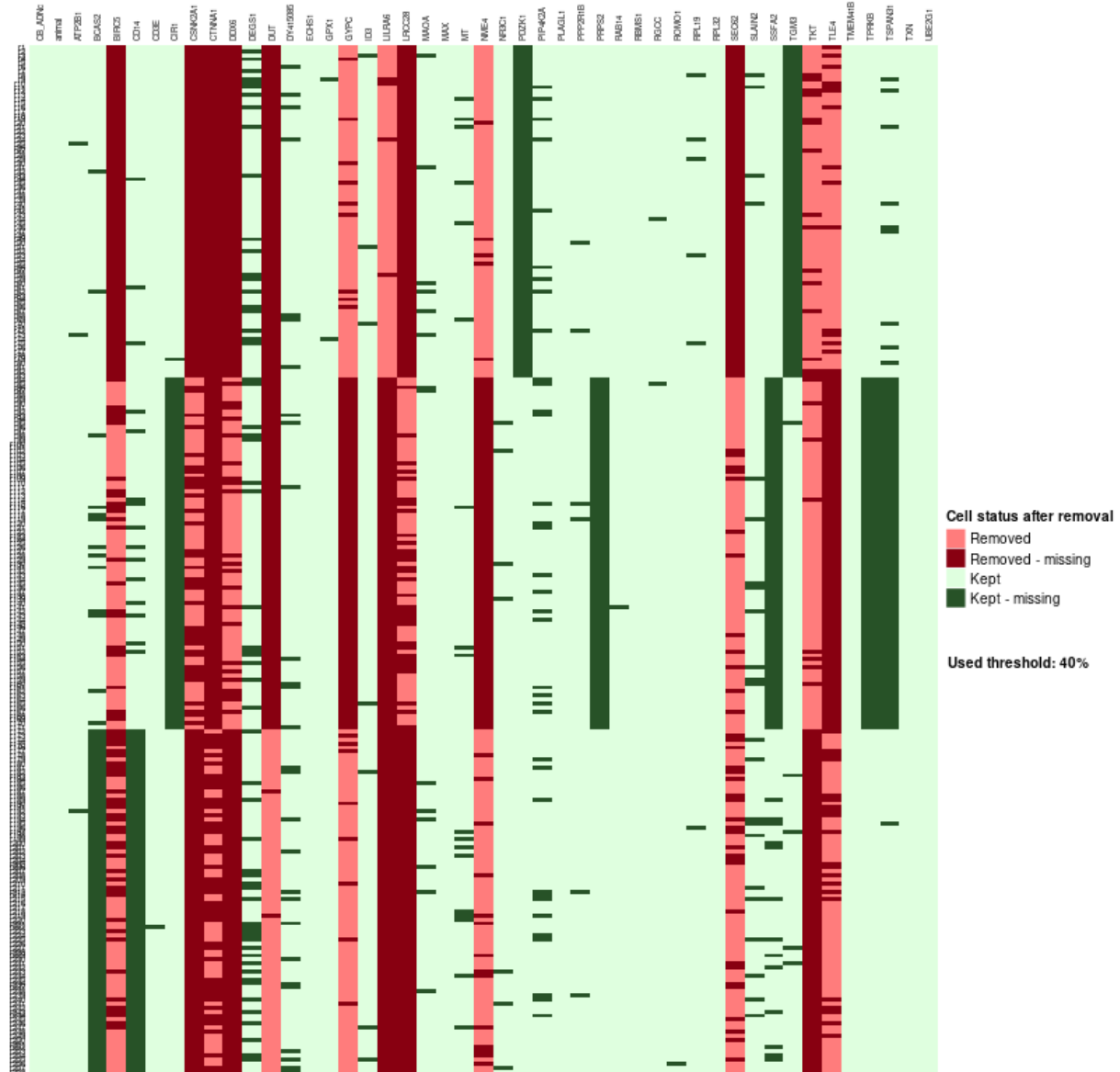
- Explore the distribution of missing values in your dataset
- **Impute** missing values with PCA, k-means or by zero.
- OR **remove** individuals / variables with the largest proportions of missing values.

How to set options?

- “Direction” is used to choose to remove either entire individuals (rows) or entire variables (columns).
- The chosen percentage corresponds to the maximum acceptable missingness. For instance, if 30% is chosen, it means that all individuals or variables with more than 30% of missing values will be removed.

Check help ? for further advice.

qPCR-1: original dataset showing variable that will be removed




Missing values

[Explore missing values](#)
[Remove missing values](#)
[Impute missing values](#)

1. Select the dataset to analyze ?

Select a dataset

 PICLET_qPCR1P218 ▾

Imputation method

PCA ▾

Type of variables to impute

Auto ▾

 Run

2. Plots and numerical summaries

Purpose of Missing values:

Handle missing values in your datasets

- Explore the distribution of missing values in your dataset
- **Impute** missing values with PCA, k-means or by zero.
- OR **remove** individuals / variables with the largest proportions of missing values.

How to set options?

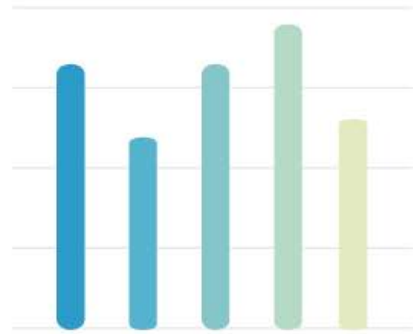
Imputation method can be chosen according to what you know on the missing values and subsequent analyses:

- **imputation by zeros** is dedicated to cases where missing values are due to a measurements below the detection threshold. It is a very basic approach to address this problem but certainly not the best;
- **imputation by PCA** is well designed when you want to use PCA, MFA, or PLS (for instance) afterwards because it best preserves the projection of your individuals on PC axes;
- **imputation by k-nearest neighbors** is based on the idea that two individuals that are similar on observed values also have similar values for unobserved variables. It best preserves the distances between individuals and is well adapted prior clustering.

The last two methods are only valid when data are missing at random.

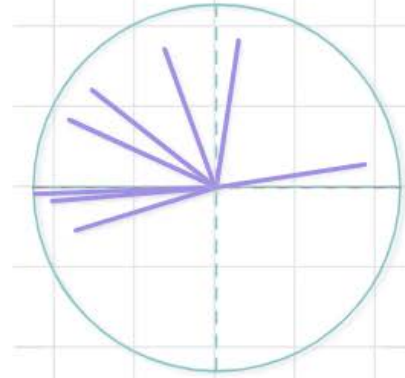
In addition, for PCA and KNN you can choose to impute **only certain types of variables** (only numerical or only categorical variables). Setting this option to "Auto" imputes only variables of the most present type while "Mixed" imputes both numerical and categorical variables.

Let's explore a dataset!



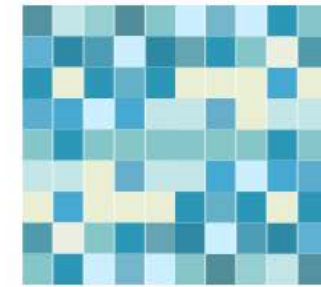
Explore variables in a dataset

Obtain numerical summaries and plots for a few variables.



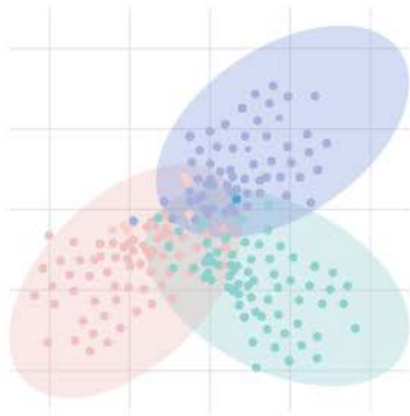
Explore a dataset with PCA

Perform Principal Component Analysis on a dataset.



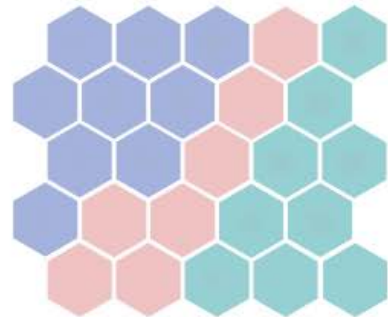
Explore a dataset with a heatmap

Obtain the heatmap of a dataset.



Clustering

Cluster the individuals of a dataset.



Self-Organizing Map

Use SOM as a clustering and visualization method.

Explore a dataset with PCA

1. Display individuals on components ?

Component to display on X (horizontal)

Component to display on Y (vertical)

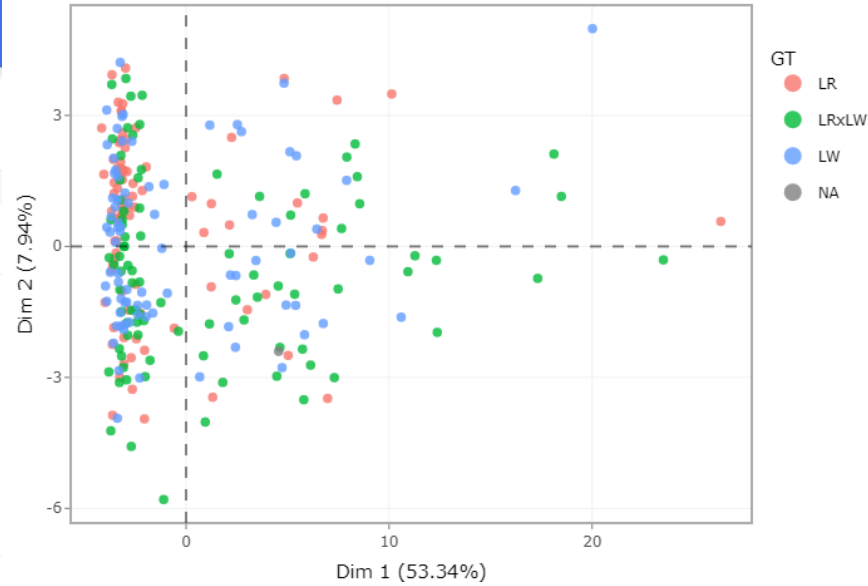
Color

Shape

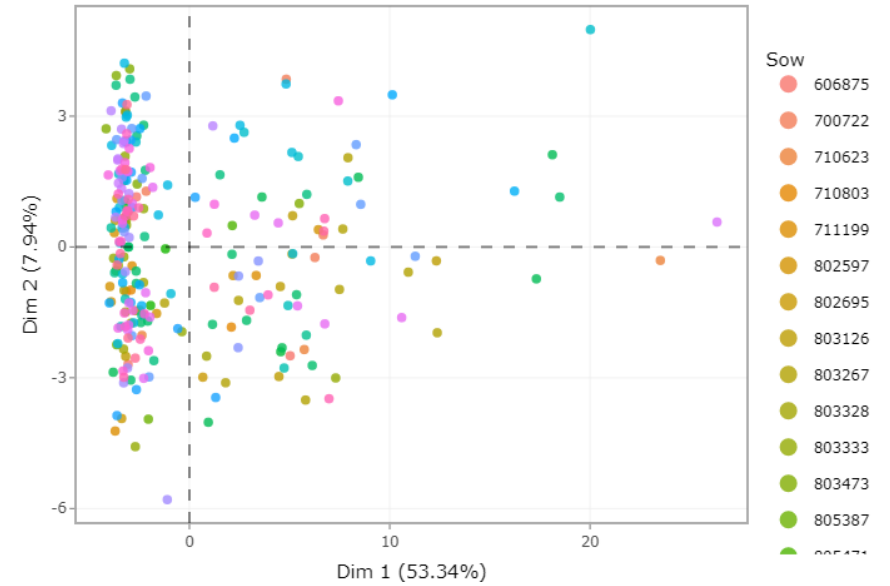
Size

Plot individuals

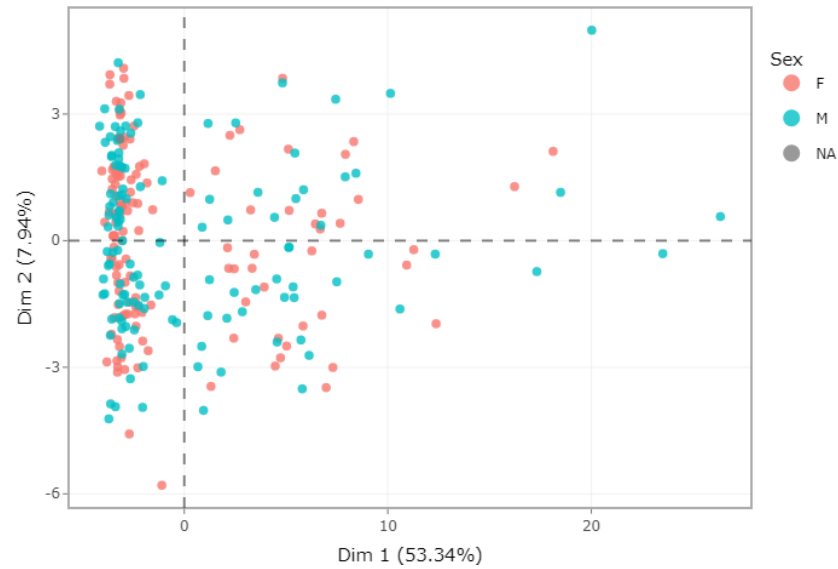
PCA on Data_qPCR_Piclet_e_Imputed_1: projection of individuals on the F



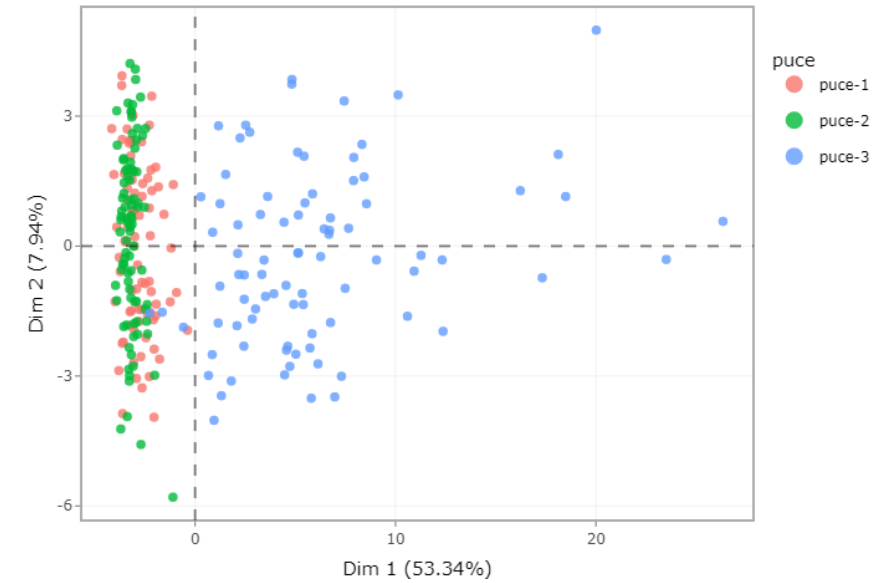
PCA on Data_qPCR_Piclet_e_Imputed_1: projection of individuals on the F



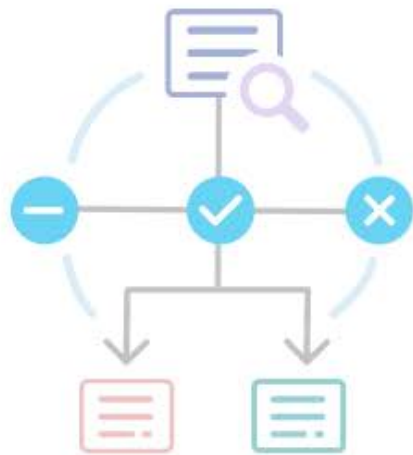
PCA on Data_qPCR_Piclet_e_Imputed_1: projection of individuals on the F



PCA on Data_qPCR_Piclet_e_Imputed_1: projection of individuals on the F

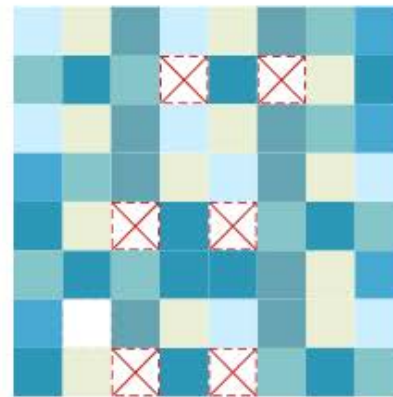


Let's edit data!



Dataset edition

Provides several ways to modify a dataset.



Missing values

Explore, remove or impute missing values on a dataset.



Normalize dataset

Correct technical biases and prepare your data for further analyses.

Normalize dataset

1. Select a dataset, choose a normalization type and set options ?

Select a dataset

 PICLET_qPCR1P218_Imputed_2 ▾

Choose normalization type

- Correct batch effect ▾
- Log-transformation
- Centering and scaling to unit variance
- Correct batch effect**
- Quantile normalization

 Run

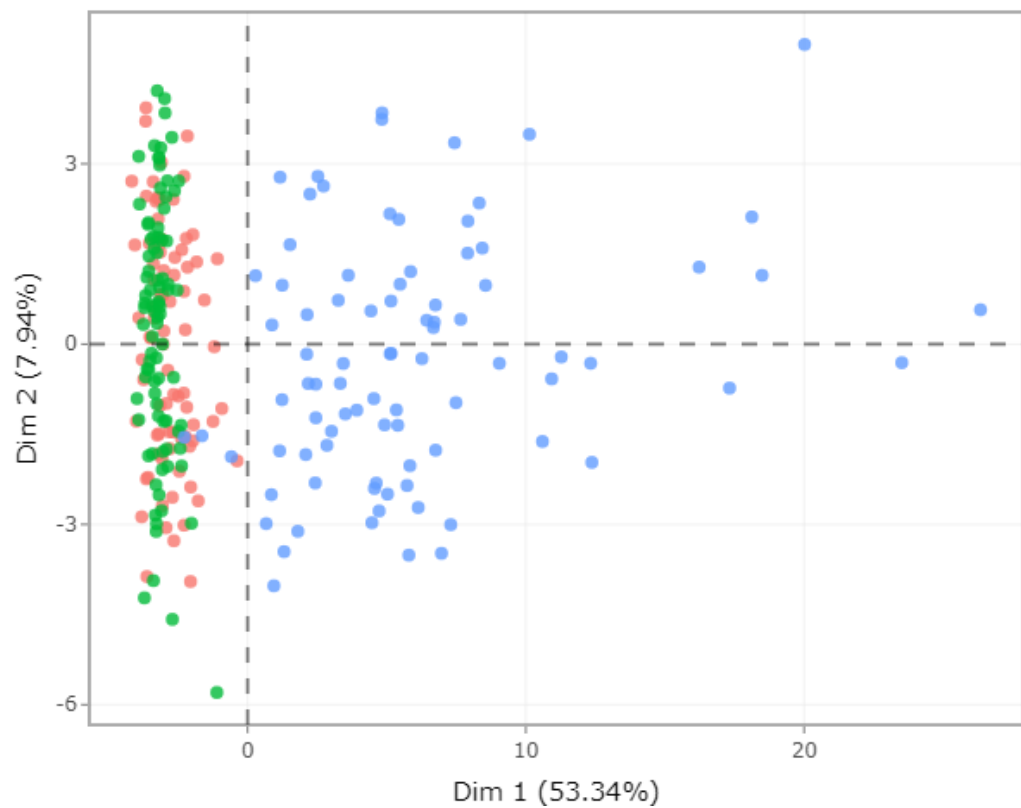
2. Dataset

Purpose of Normalize dataset:

Correct technical biases and prepare your datasets for further analyses

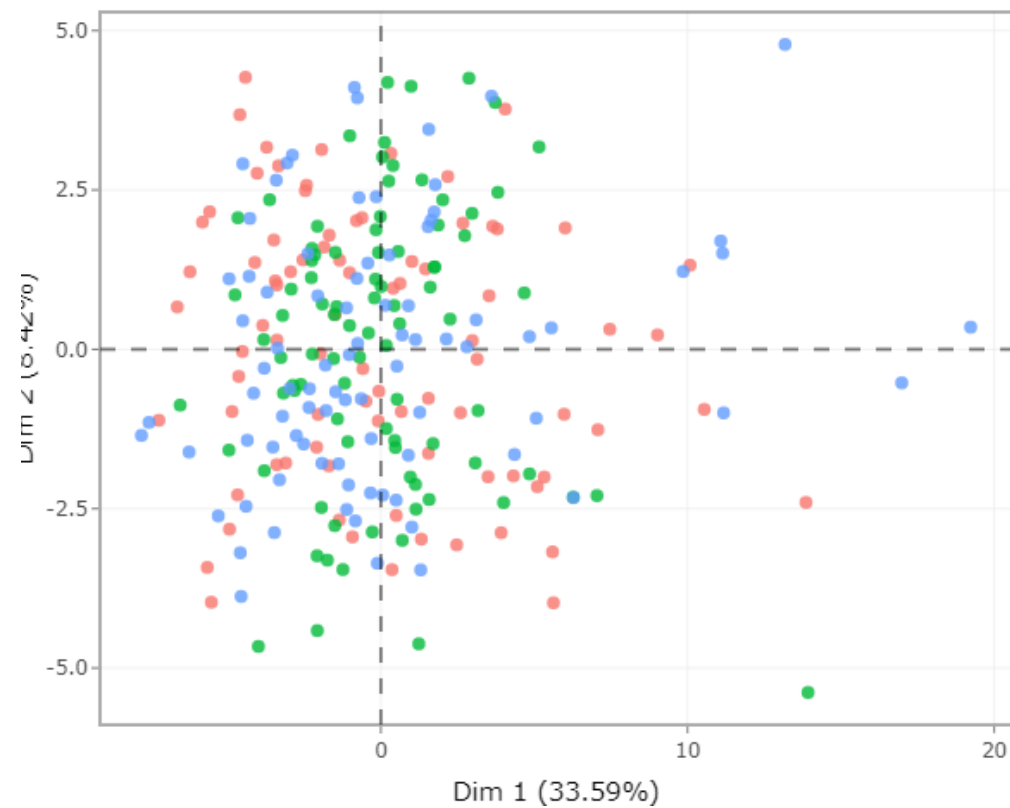
- **Basic transformations** (log, ...).
- **Normalizations specific of certain data types** (RNA-seq, ...).
- Normalization for the **correction of an explicit batch effect**.
- All normalization methods come with **diagnostic plots**.

PCA on Data_qPCR_Piclet_e_Imputed_1: projection of individuals on the F



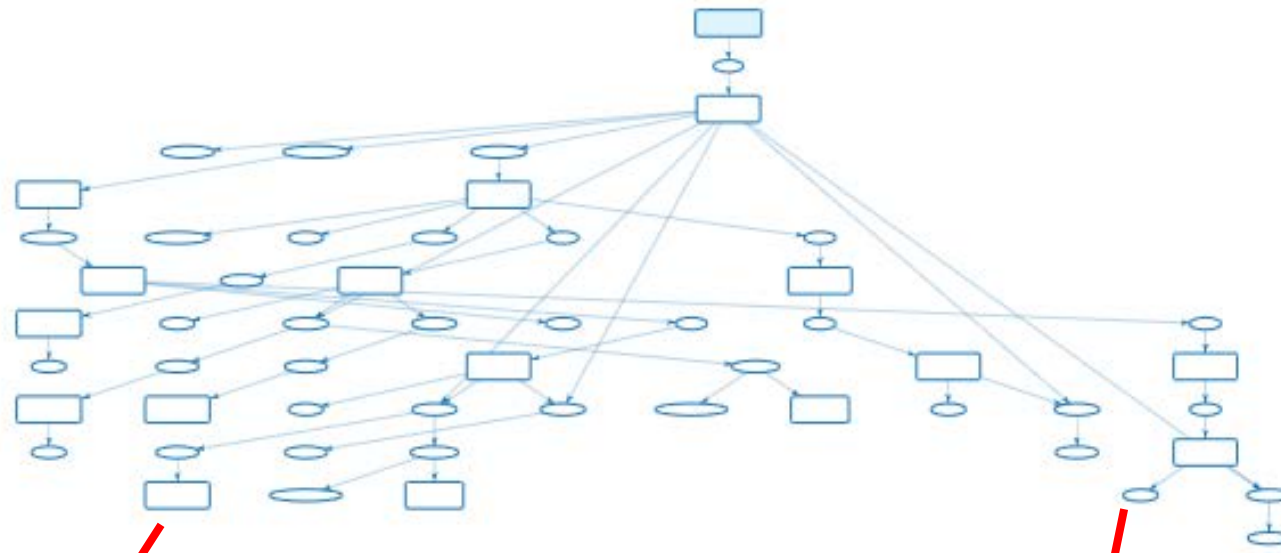
puce
● puce-1
● puce-2
● puce-3

PCA on Data_qPCR_Piclet_e_Imputed_1_normalized_1: projection of ind



puce
● puce-1
● puce-2
● puce-3

Workflow



All datasets

Remove Add

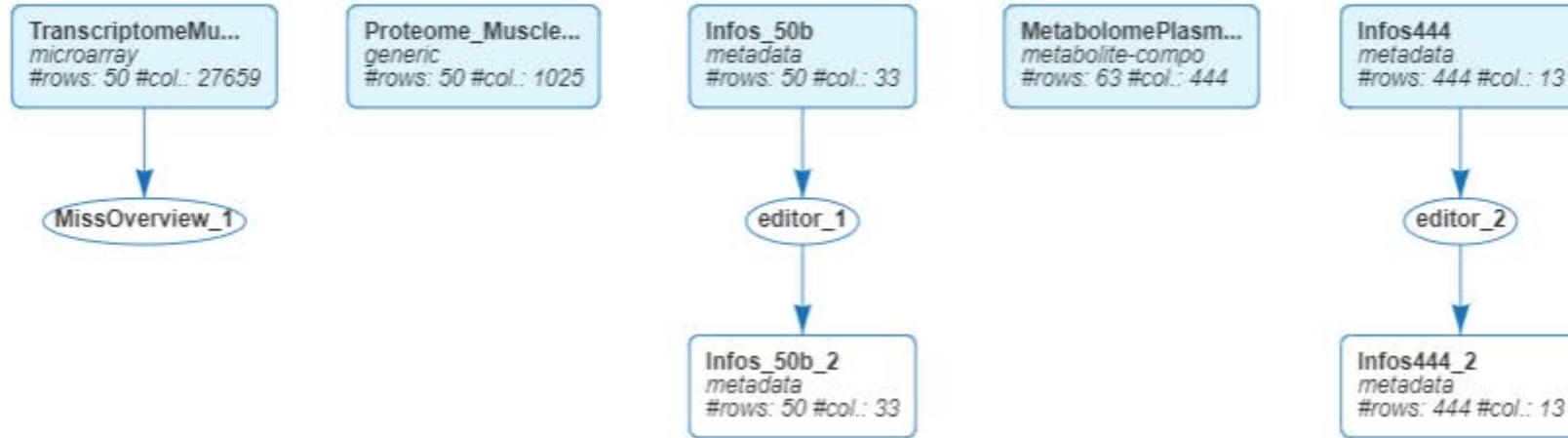
| <input type="checkbox"/> | Name | Type | # Rows | # Columns | Log | Normalized | |
|--------------------------|--------------------|---------|--------|-----------|-----|------------|--|
| <input type="checkbox"/> | PICLET_qPCR1P21... | generic | 257 | 45 | no | no | |
| <input type="checkbox"/> | PICLET_qPCR1P21... | generic | 257 | 25 | nc | nc | |
| <input type="checkbox"/> | PICLET_qPCR1P21... | generic | 257 | 45 | nc | nc | |
| <input type="checkbox"/> | PICLET_qPCR1P21... | generic | 257 | 25 | nc | nc | |

All analyses

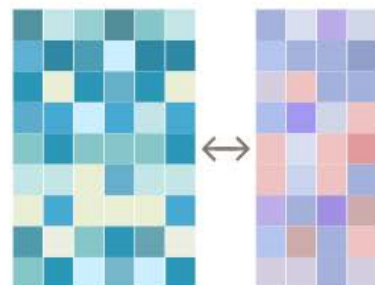
Remove Export report

| <input type="checkbox"/> | Name | Function |
|----------------------------|--------------------|----------------------|
| > <input type="checkbox"/> | editor_1 | r_edit_dataset |
| > <input type="checkbox"/> | UnivariateDatas... | r_univariate_dataset |
| > <input type="checkbox"/> | MissOverview_1 | r_missing_overview |
| > <input type="checkbox"/> | MissShowRemove_... | r_missing_showremove |
| > <input type="checkbox"/> | MissShowRemove_... | r_missing_showremove |
| > <input type="checkbox"/> | PCAobj_1 | r_famd |
| > <input type="checkbox"/> | imputeMissing_1 | r_missing_impute |
| > <input type="checkbox"/> | imputeMissing_2 | r_missing_impute |

Workflow

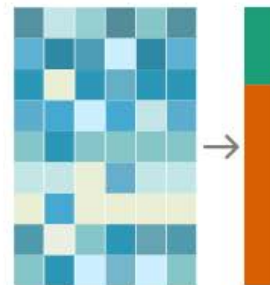


Let's integrate data!



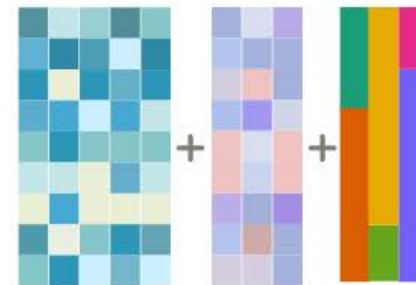
Integrate two datasets with PLS

Performs Partial Least Squares analysis on two datasets.



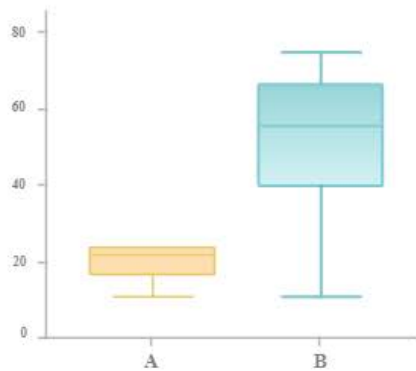
Integrate two datasets with PLS-DA

Performs Partial Least Squares analysis on two datasets.



Integrate datasets with MFA

Perform Multiple Factor Analysis on several datasets.



Differential analysis

Perform differential analysis for all numeric variables of a dataset.

Integrate two datasets with PLS


[Preprocessing](#)
[Run PLS](#)
[Explore individuals](#)
[Explore variables](#)
[Extract new data](#)

1. Select the datasets to integrate ?

Datasets

Proteome_Muscle_50 ×
Infos_50_edited ×


2. Plots and summaries

 [General information](#)
 [Venn diagram](#)
 [Upset plot](#)

Description of datasets before filtering common individuals (rows)

| Name | # rows | # col. | # missing | % missing | # numeric | # cat. | # logic | # others |
|--------------------|--------|--------|-----------|-----------|-----------|--------|---------|----------|
| Infos_50_edited | 50 | 32 | 0 | 0 | 26 | 6 | 0 | 0 |
| Proteome_Muscle... | 50 | 1025 | 0 | 0 | 1025 | 0 | 0 | 0 |

Description of datasets after filtering common individuals (rows)

| Name | # rows | # col. | # missing | % missing | # numeric | # cat. | # logic | # others |
|--------------------|--------|--------|-----------|-----------|-----------|--------|---------|----------|
| Infos_50_edited | 50 | 32 | 0 | 0 | 26 | 6 | 0 | 0 |
| Proteome_Muscle... | 50 | 1025 | 0 | 0 | 1025 | 0 | 0 | 0 |

Integrate two datasets with PLS

[Preprocessing](#)[Run PLS](#)[Explore individuals](#)[Explore variables](#)[Extract new data](#)

1. Display individuals on components ?

Component to display on X (horizontal)

Component to display on Y (vertical)

Color

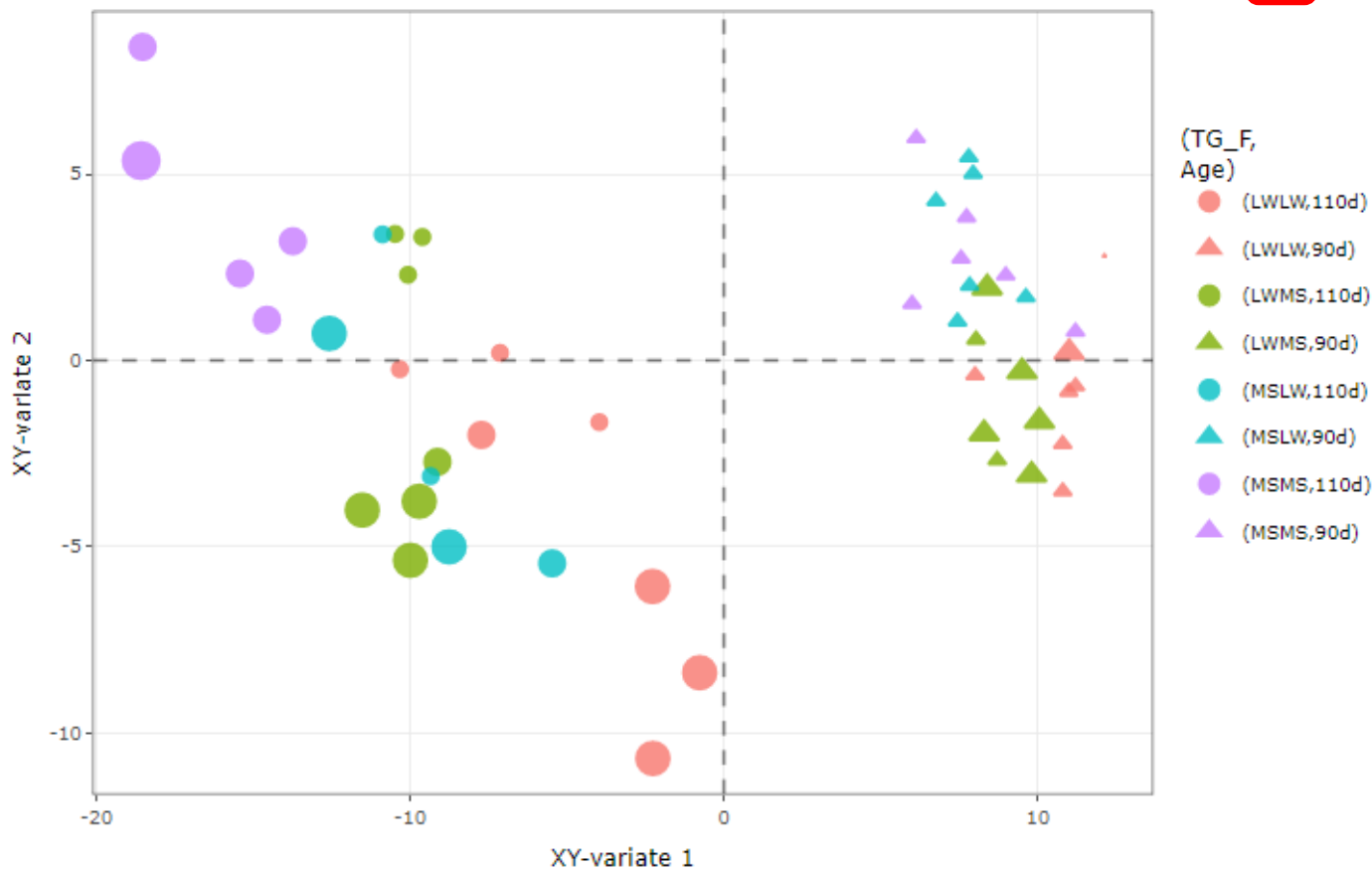
Shape

Size

[Plot individuals](#)

2. Plot individuals on components

PLS on Info_50b_e, Proteome_Muscle_50: XY-space

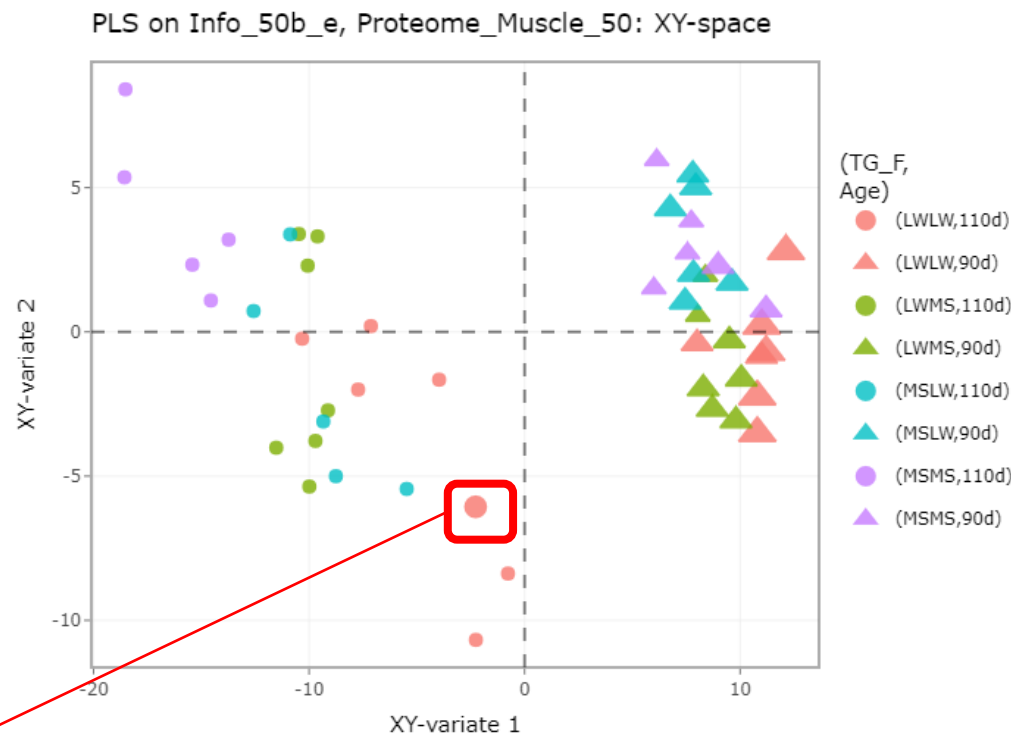
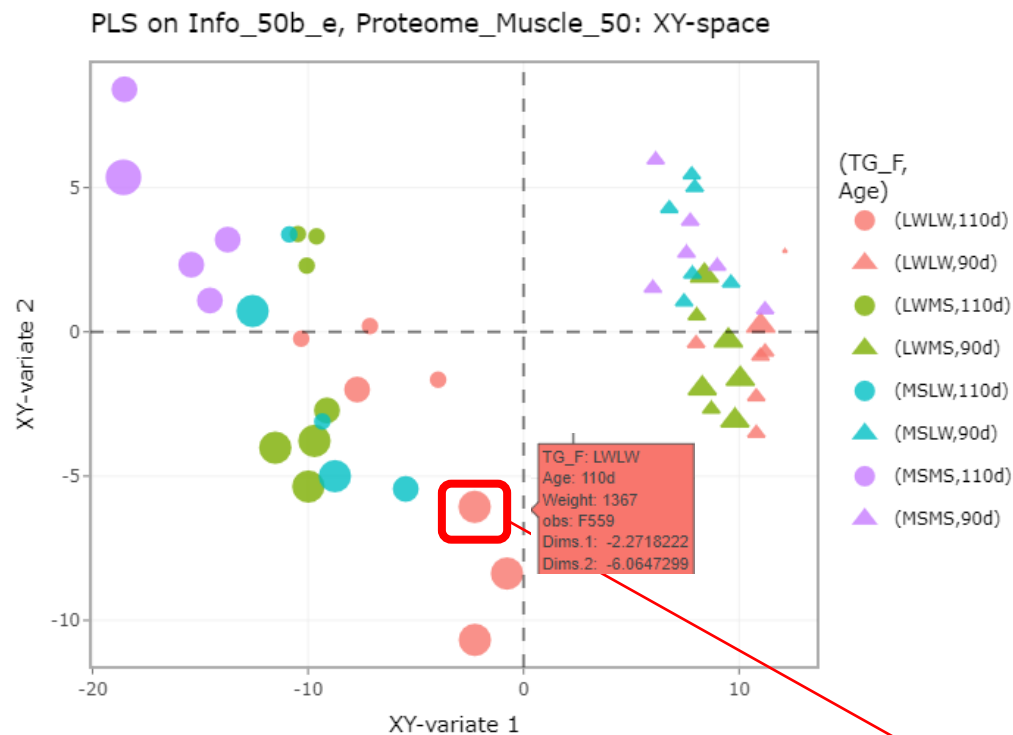
[Add to report](#)

Integrate two datasets with PLS

[Preprocessing](#)
[Run PLS](#)
[Explore individuals](#)
[Explore variables](#)
[Extract new data](#)

Size = weight

Size = MHC_E



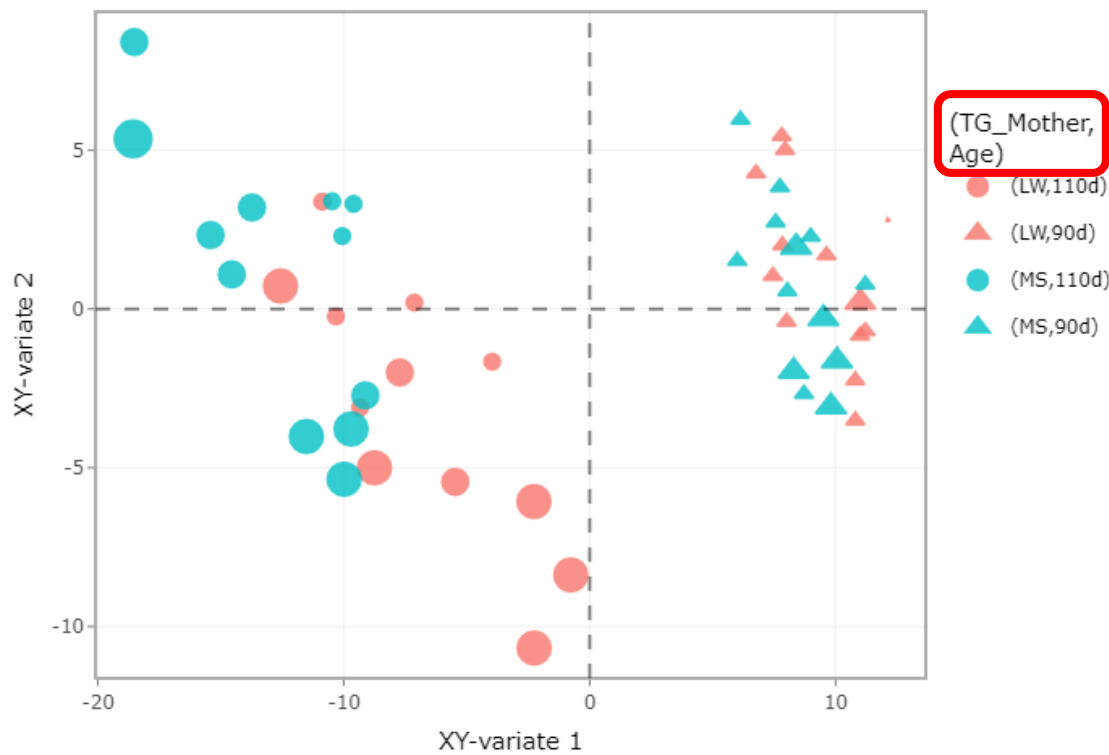
Heavy but developmental delay

Integrate two datasets with PLS

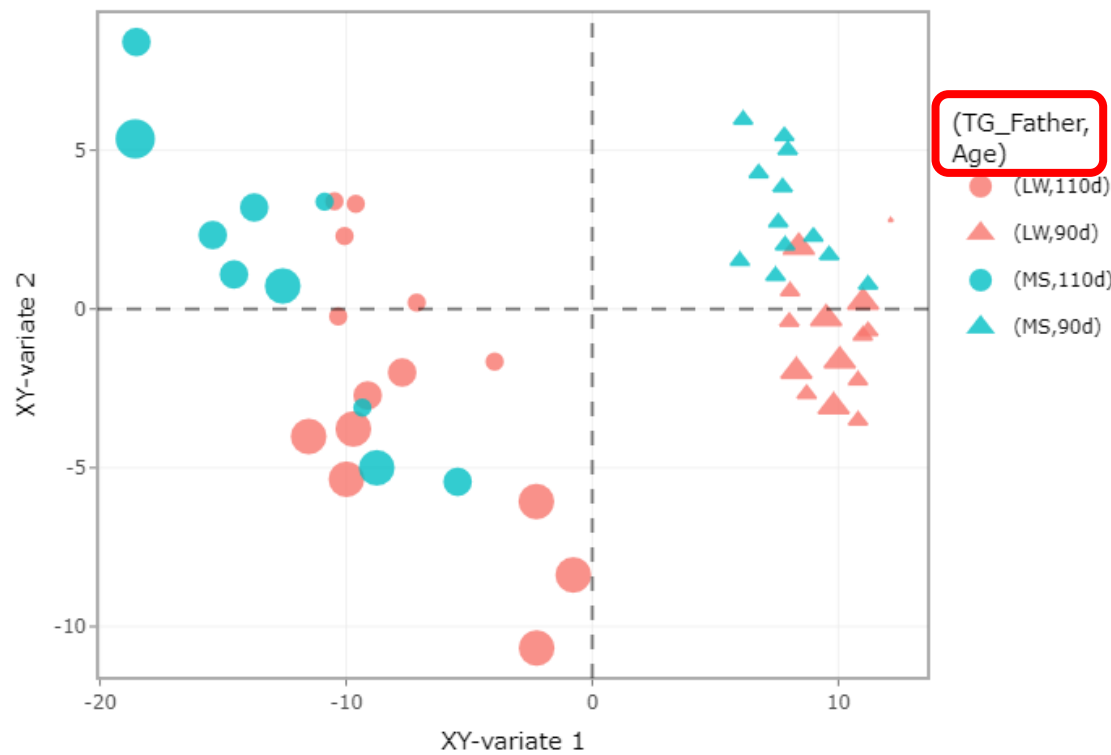
[Preprocessing](#)
[Run PLS](#)
[Explore individuals](#)
[Explore variables](#)
[Extract new data](#)

Size = weight

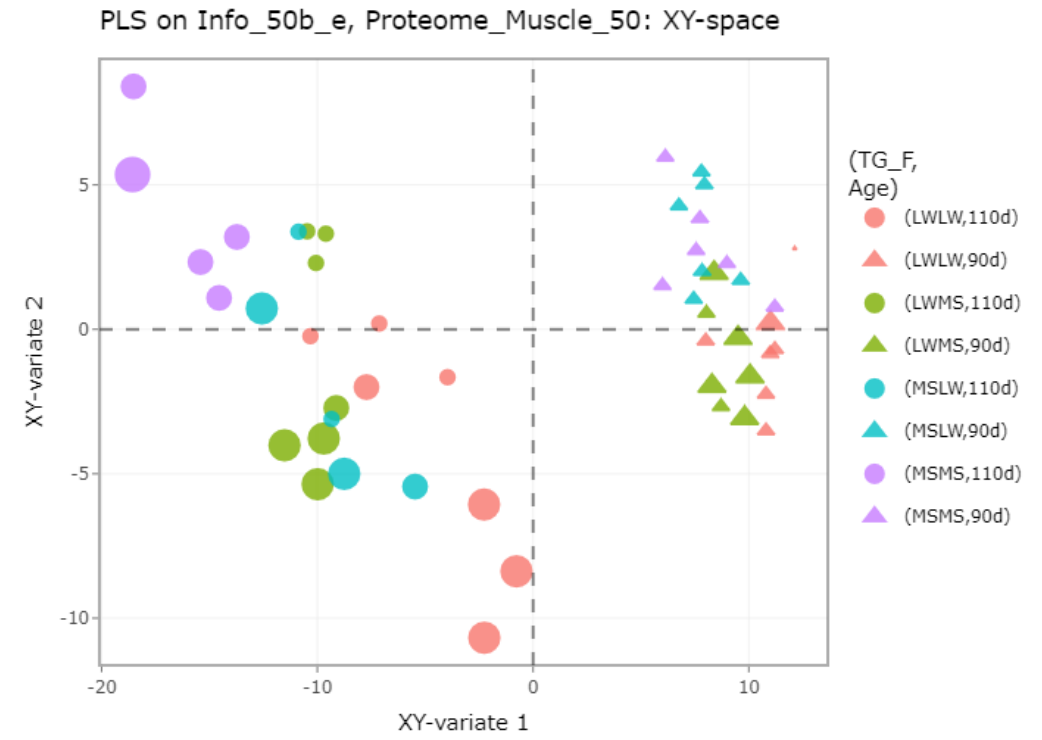
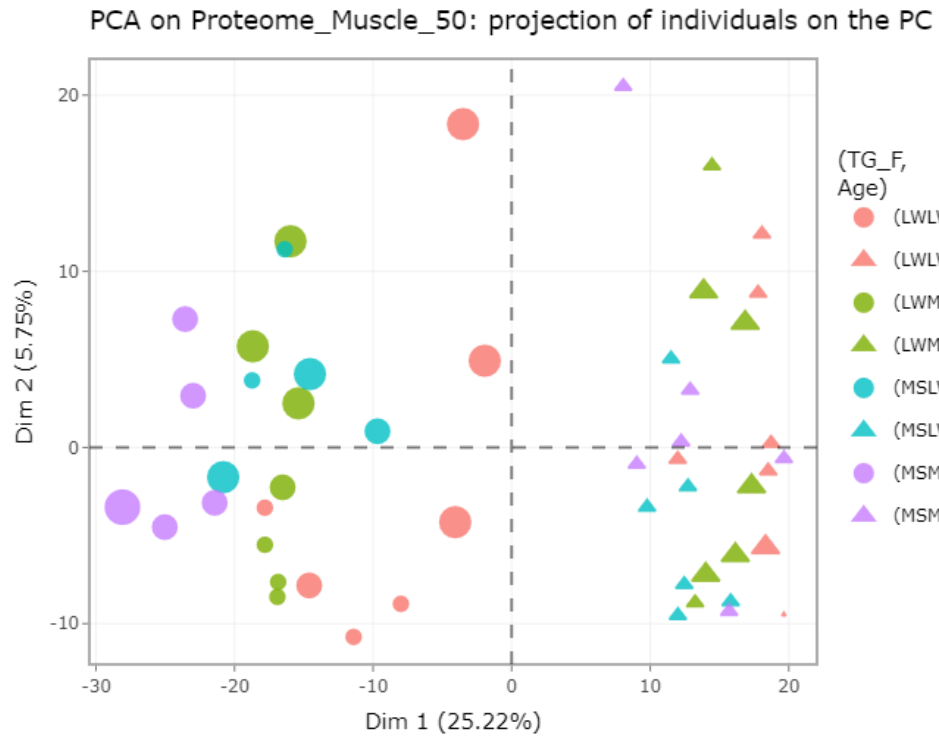
PLS on Info_50b_e, Proteome_Muscle_50: XY-space



PLS on Info_50b_e, Proteome_Muscle_50: XY-space



PCA vs PLS



Size = weight

Integrate two datasets with PLS

Preprocessing

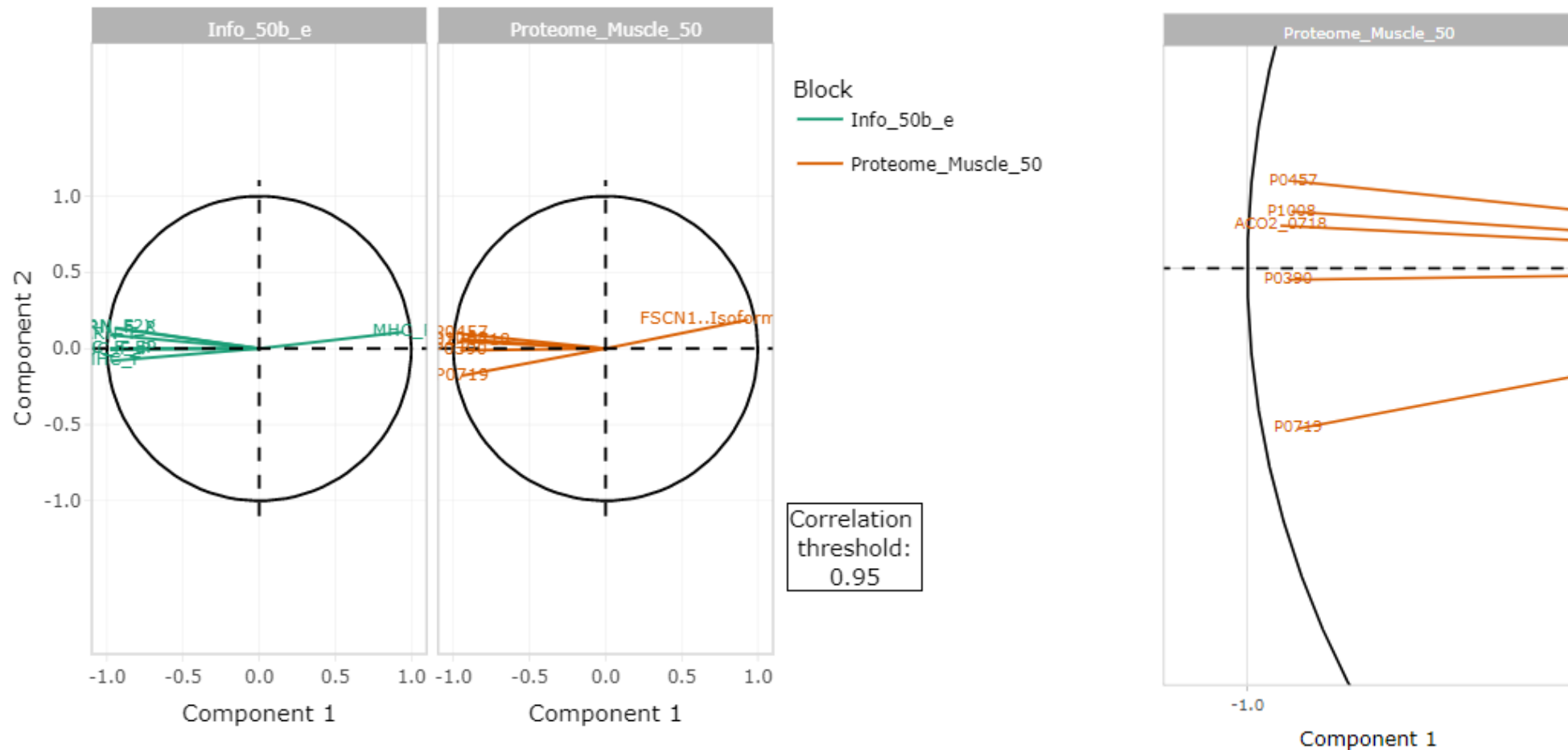
Run PLS

Explore individuals

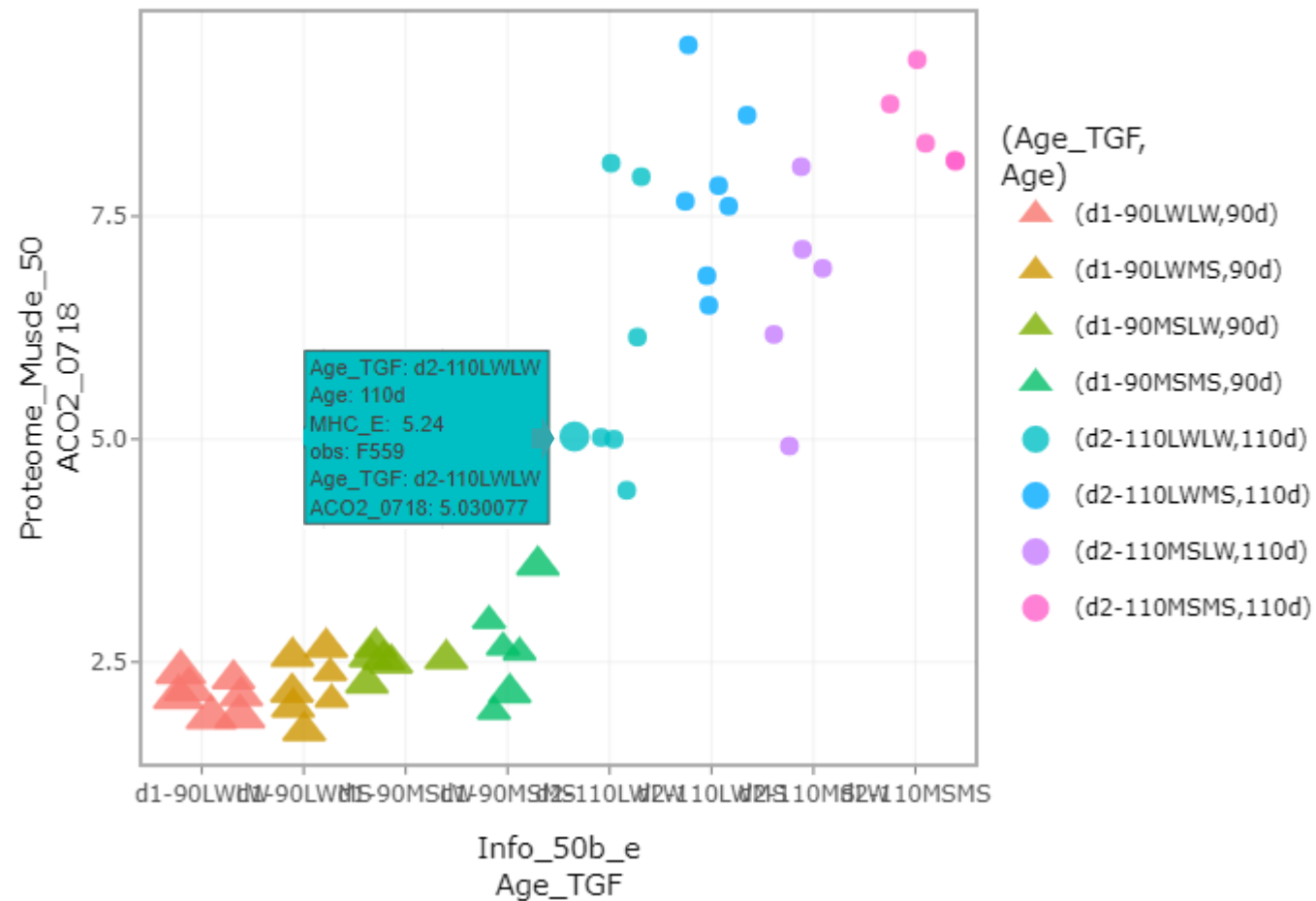
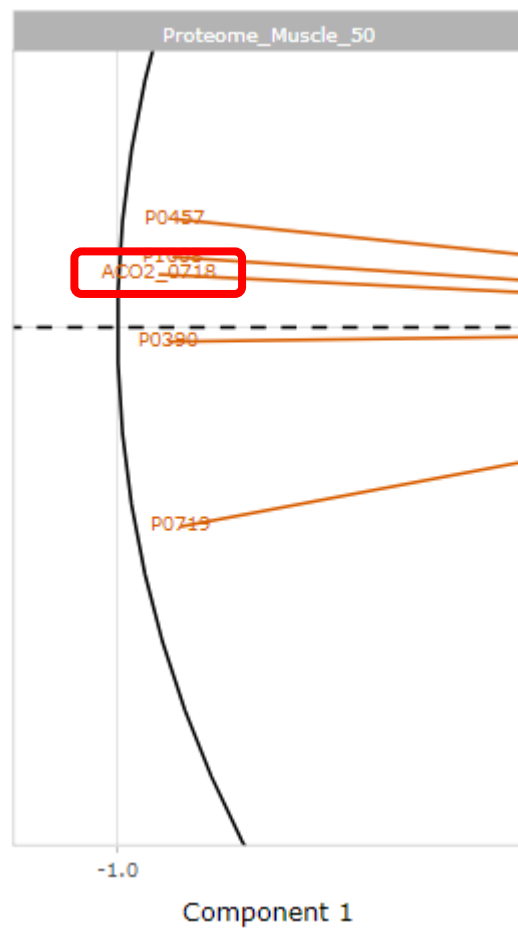
Explore variables

Extract new data

PLS on Info_50b_e, Proteome_Muscle_50: correlations of variables with



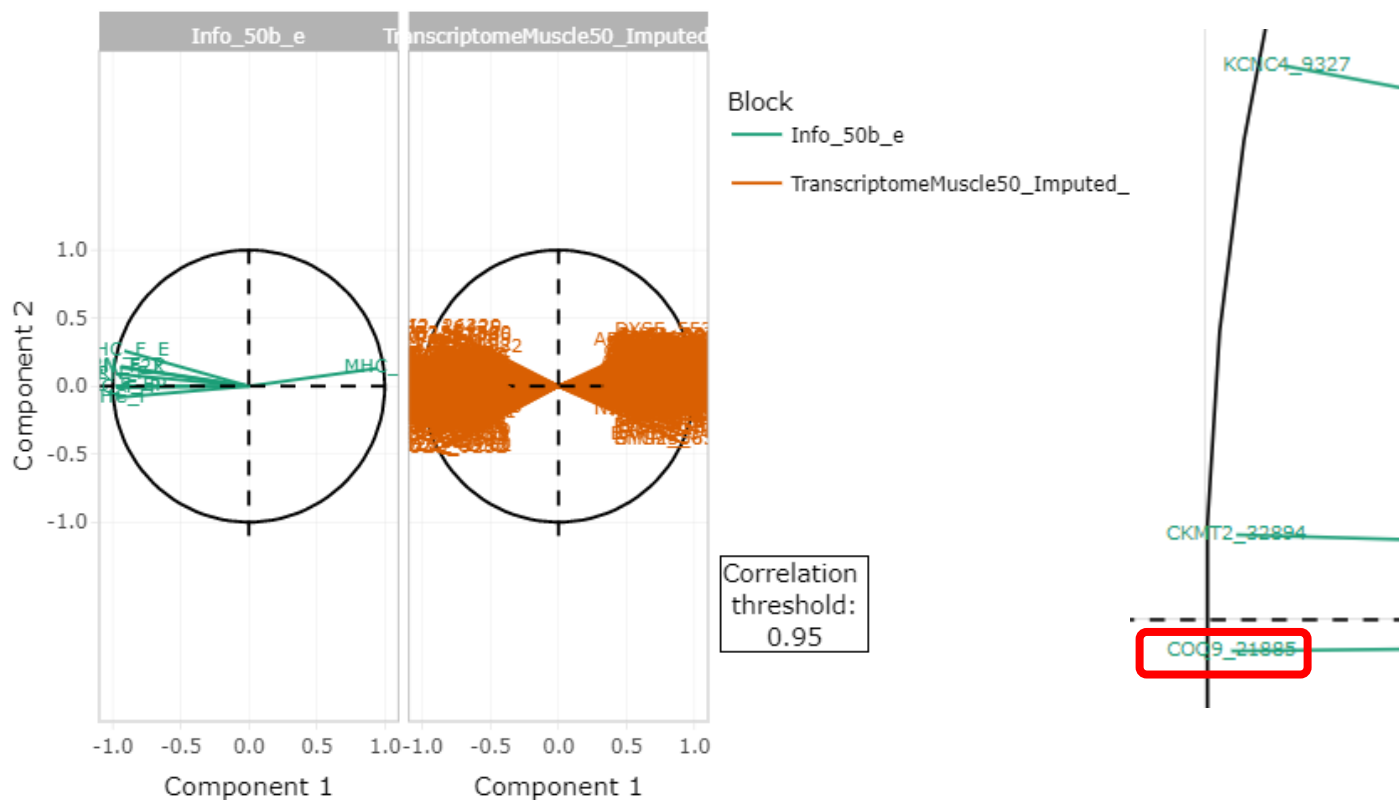
Size = MHC_E



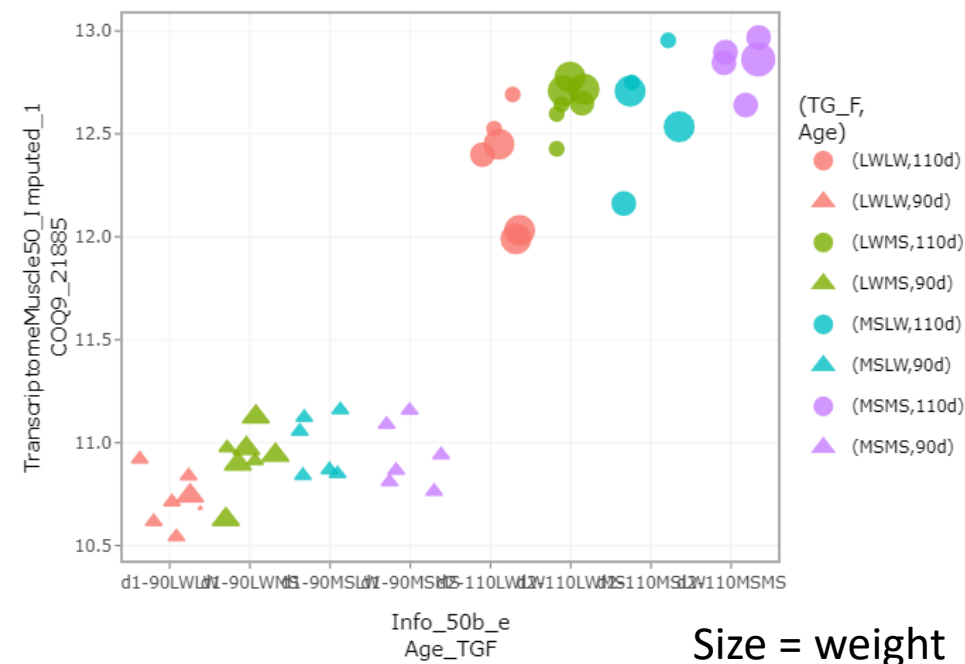
Integrate two datasets with PLS

[Preprocessing](#)
[Run PLS](#)
[Explore individuals](#)
[Explore variables](#)
[Extract new data](#)

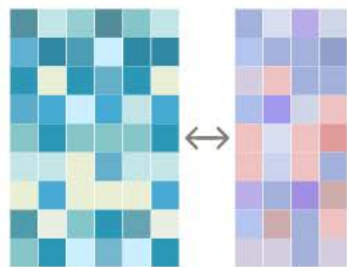
PLS on Info_50b_e, TranscriptomeMuscle50_Imputed_1: correlations of



Explore variables in a dataset

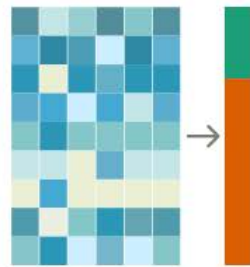


Let's integrate data!



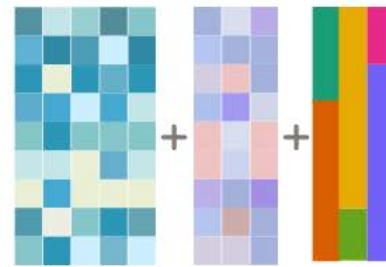
Integrate two datasets with PLS

Performs Partial Least Squares analysis on two datasets.



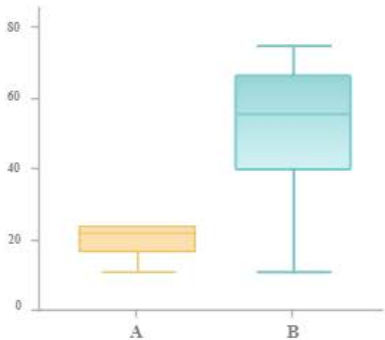
Integrate two datasets with PLS-DA

Performs Partial Least Squares analysis on two datasets.



Integrate datasets with MFA

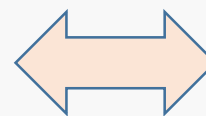
Perform Multiple Factor Analysis on several datasets.



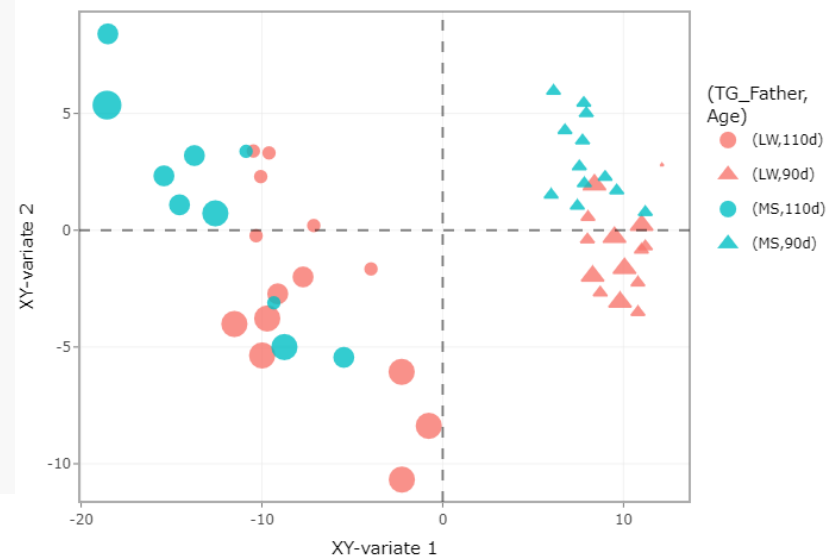
Differential analysis

Perform differential analysis for all numeric variables of a dataset.

It is easy to discriminate the age but what about the genotype?



PLS on Info_50b_e, Proteome_Muscle_50: XY-space



Integrate two datasets with PLS-DA

[Preprocessing](#)
[Run PLS-DA](#)
[Explore individuals](#)
[Explore variables](#)
[Extract new data](#)

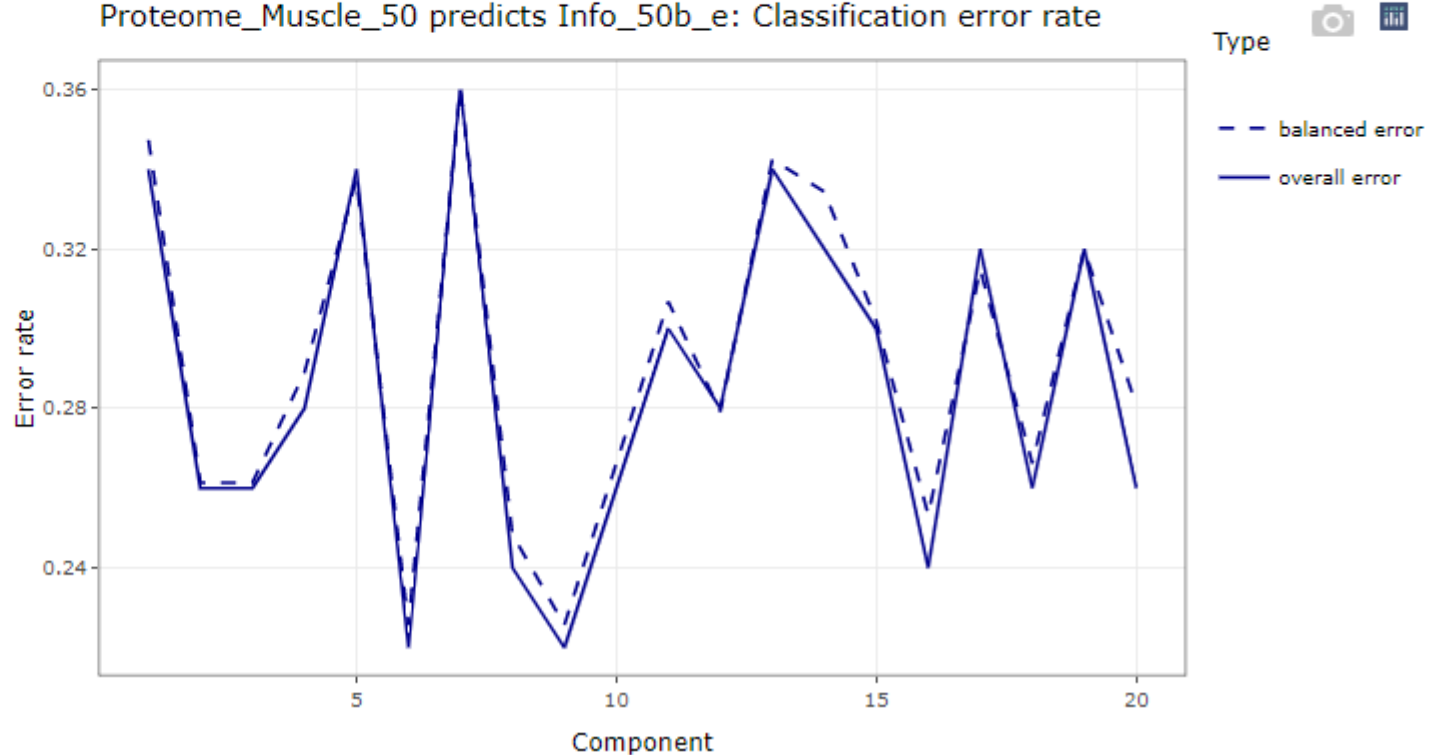
1. Run PLS-DA ?

Run

TG_Father
2 levels

2. Plots

Proteome_Muscle_50 predicts Info_50b_e: Classification error rate



Integrate two datasets with PLS-DA

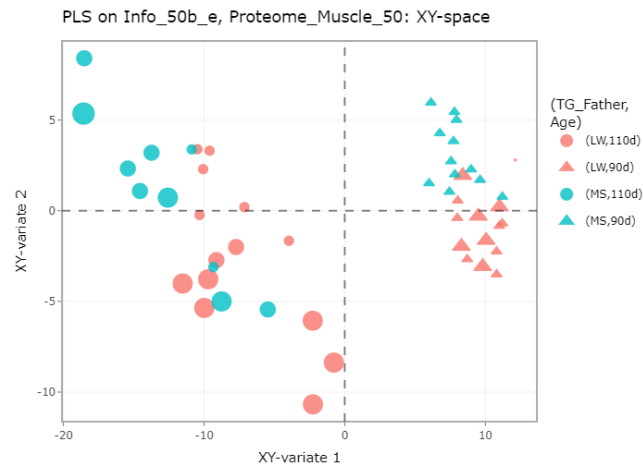
Preprocessing

Run PLS-DA

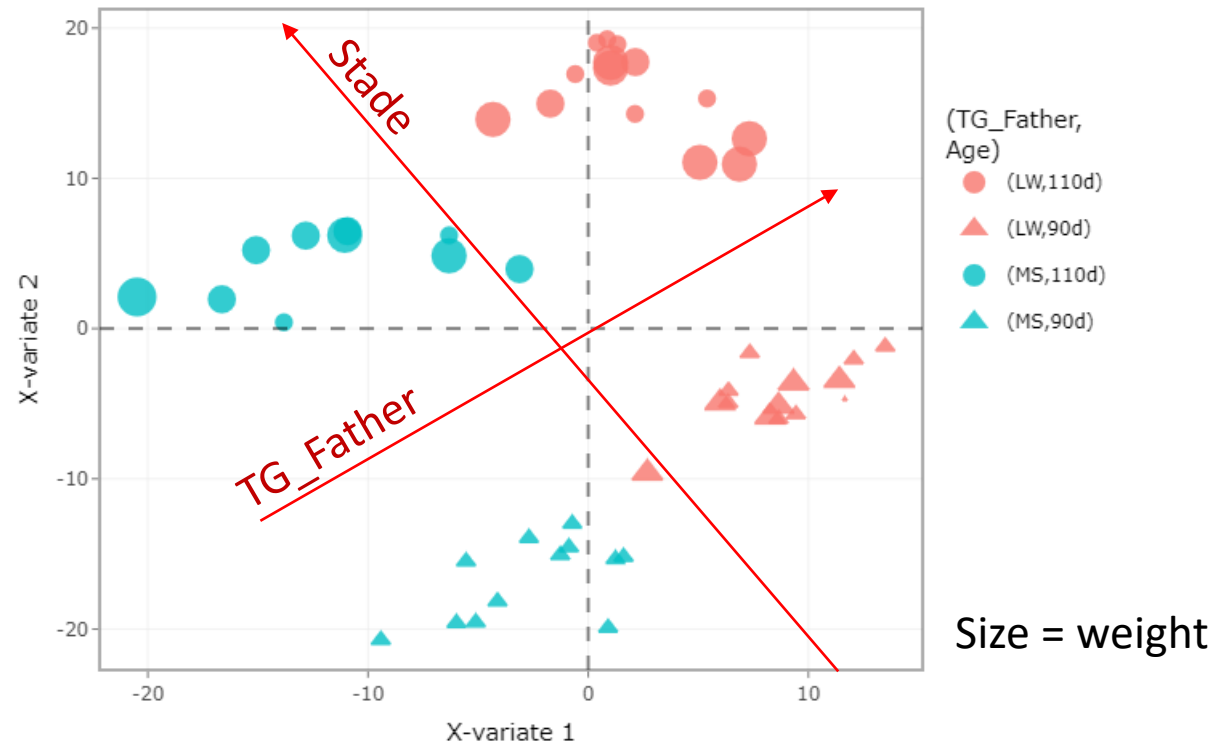
Explore individuals

Explore variables

Extract new data



PLS-DA on Proteome_Muscle_50, Info_50b_e: X-space



Integrate two datasets with PLS-DA

Preprocessing

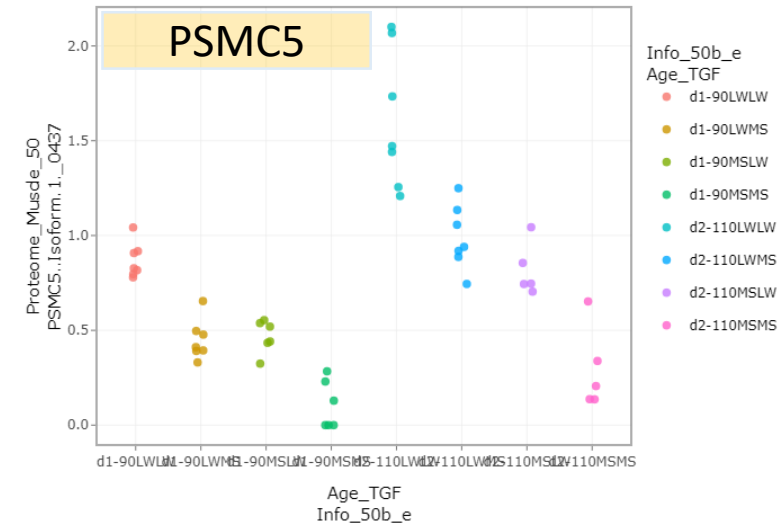
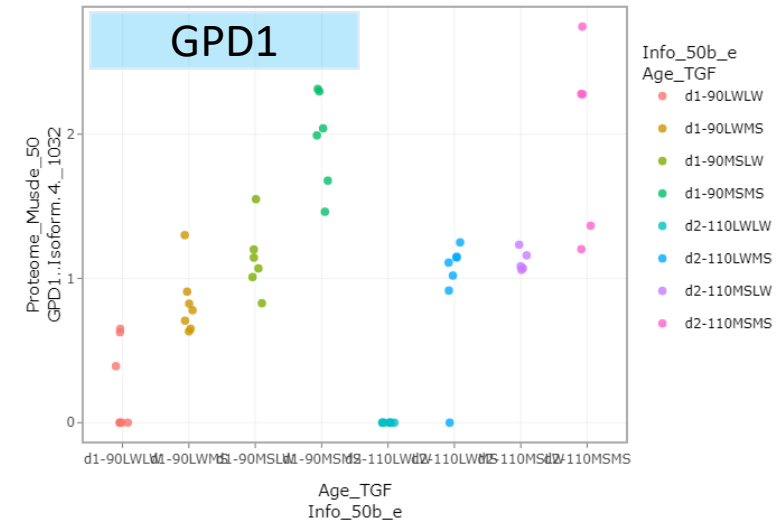
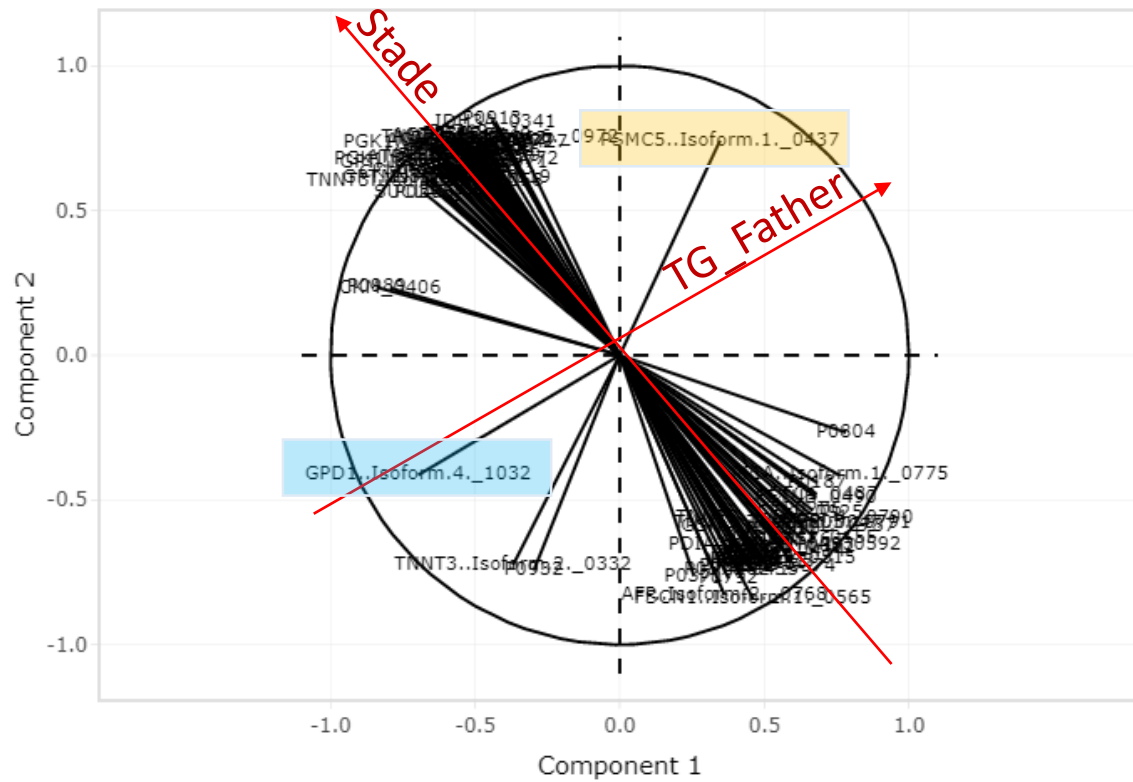
Run PLS-DA

Explore individuals

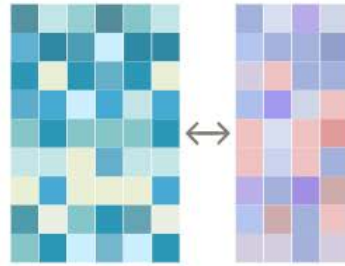
Explore variables

Extract new data

PLSDA on Proteome_Muscle_50, Info_50b_e: correlations of variables with

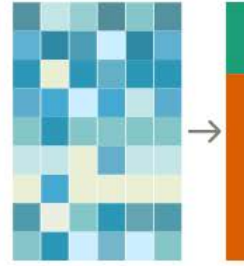


Let's integrate data!



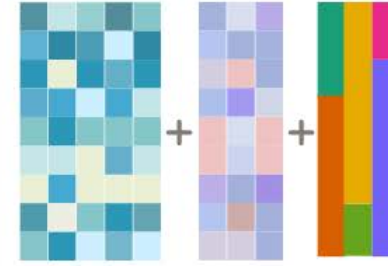
Integrate two datasets with PLS

Performs Partial Least Squares analysis on two datasets.



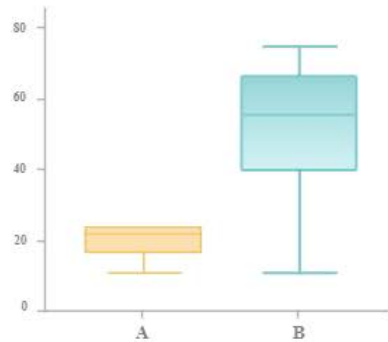
Integrate two datasets with PLS-DA

Performs Partial Least Squares analysis on two datasets.



Integrate datasets with MFA

Perform Multiple Factor Analysis on several datasets.



Differential analysis

Perform differential analysis for all numeric variables of a dataset.

Differential analysis

[Preprocessing](#)
[Multiple tests](#)
[Posthoc tests](#)
[Extract dataset](#)

1. Run multiple tests ?

Type

parametric
 non-parametric
 automatic

Correction

BH ▼

Threshold

0.05 ✓

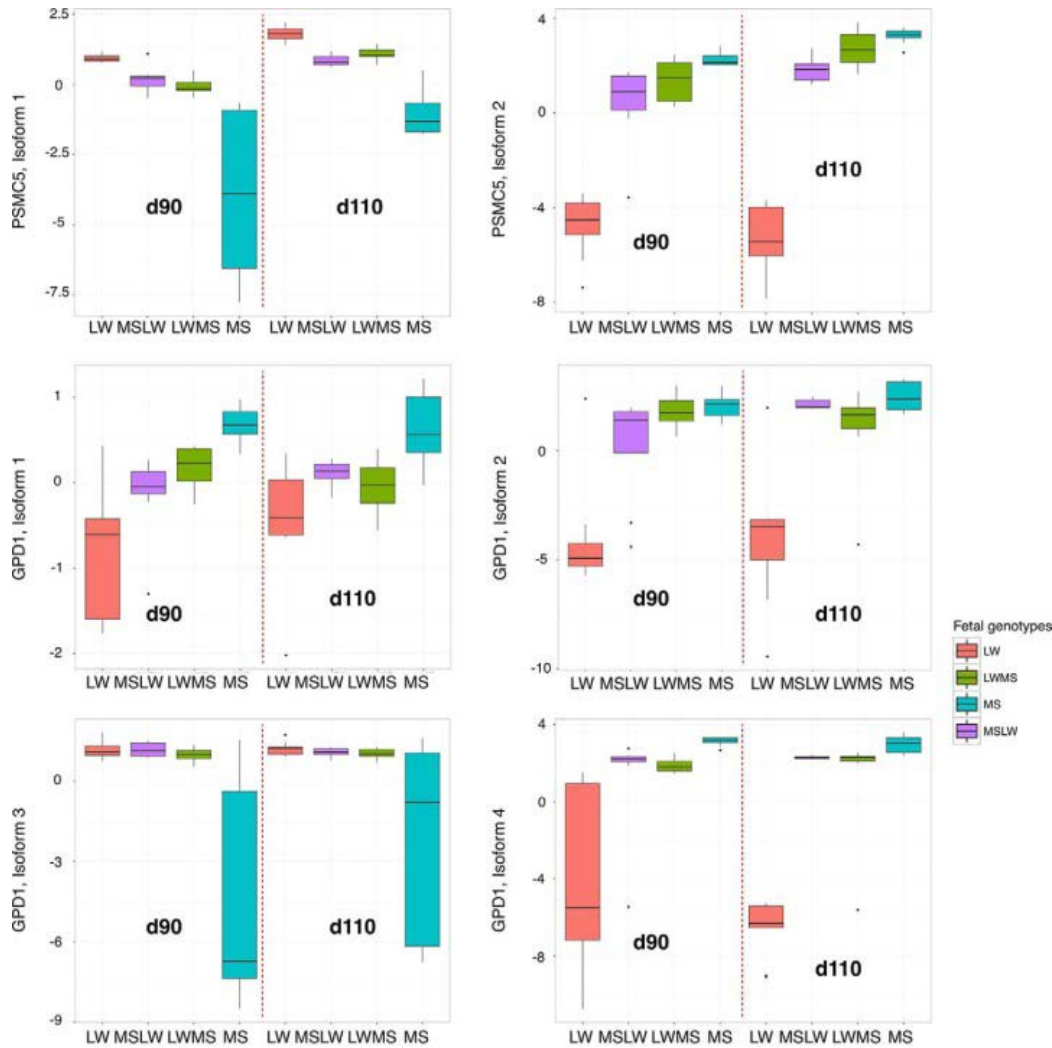
Run

2. Explore results of multiple tests

[Plots](#)
[Summary](#)

Test results (Kruskal-Wallis test, corrected with 'BH') for Proteome_Muscle_50 vs Info_50b_e **TG_Father**

| Variables | Test stats. | p-values | Adj. p-values | Signs |
|------------------------|-------------|--------------|---------------|-------|
| 28 proteins BH<0.05 | | | | |
| GPD1..Isoform.4._1032 | 26.5851 | 2.5218e-7 | 0.0003 | 1 |
| LXN_0198 | 20.745 | 0.000005247 | 0.0021 | -1 |
| P0987 | 20.4733 | 0.0000060468 | 0.0021 | -1 |
| TNNT3..Isoform.3._0355 | 18.995 | 0 | 0.0022 | 1 |
| PSMC5..Isoform.1._0437 | 18.9987 | 0 | 0.0022 | -1 |
| P0988 | 19.4156 | 0 | 0.0022 | 1 |
| TF_0852 | 18.3197 | 0 | 0.0027 | -1 |



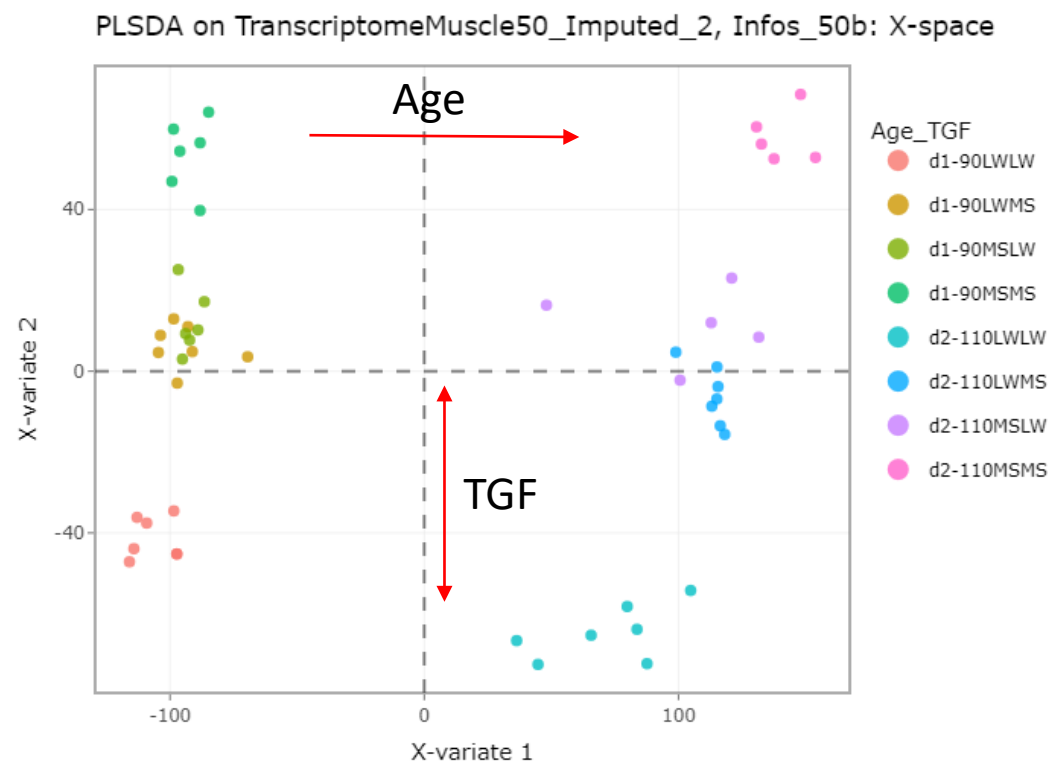
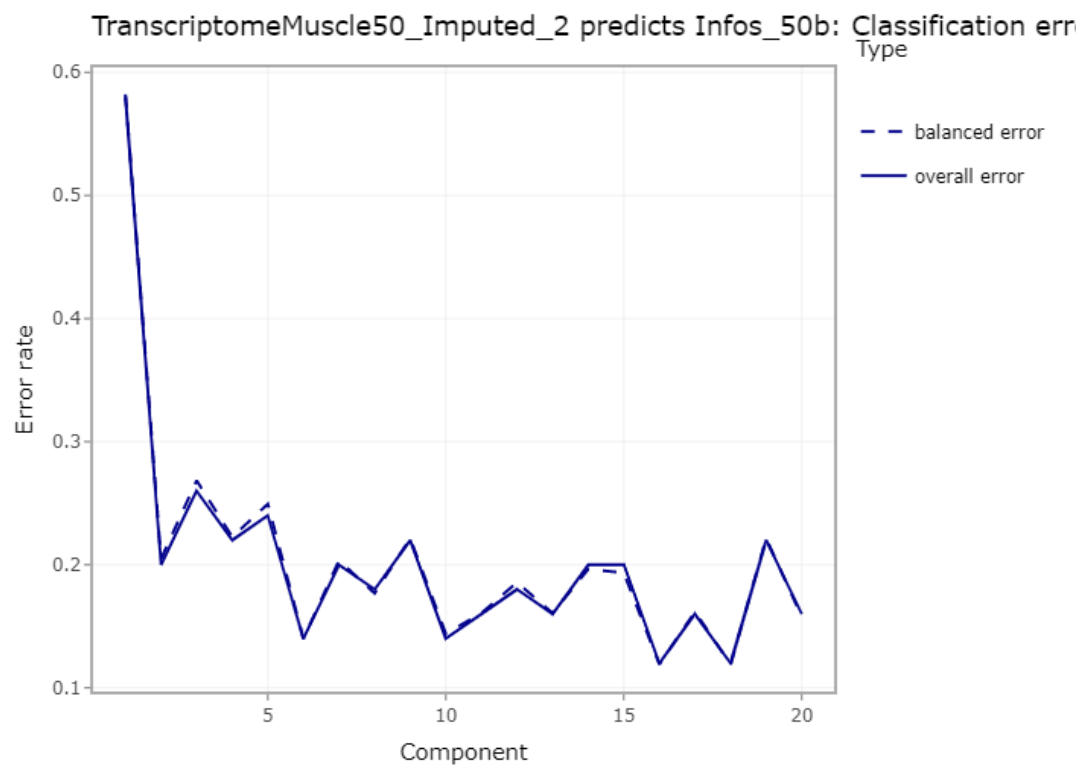
PSMC5,
 Proteasome 26S Regulatory Subunit, ATP-dependent
 degradation of ubiquitinated proteins, 5
 2 isoforms

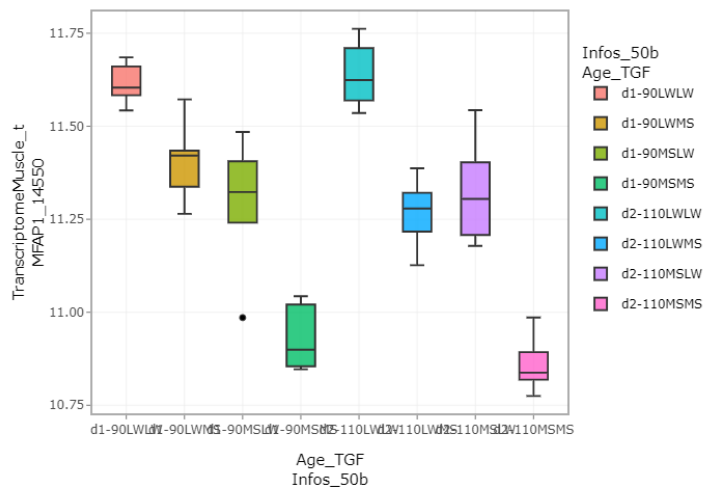
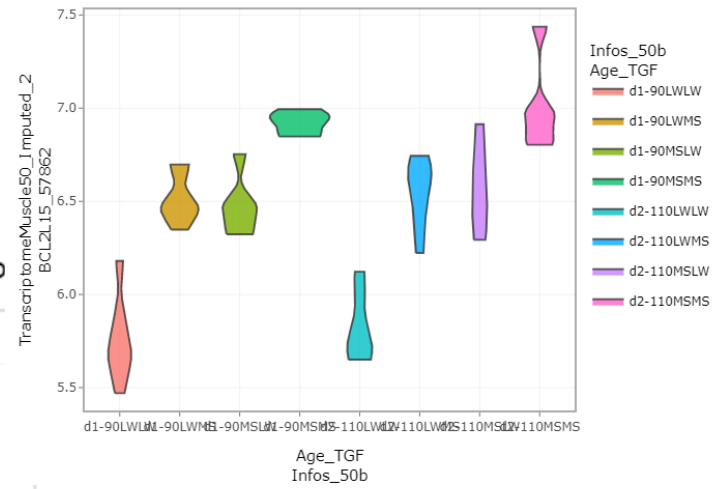
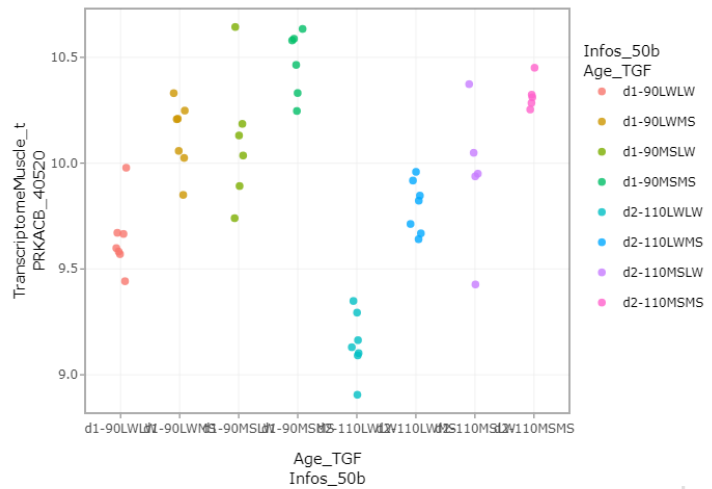
PGD1
 Glycerol-3-phosphate dehydrogenase 1, cytoplasmic
 4 isoforms

Integrate two datasets with PLS-DA

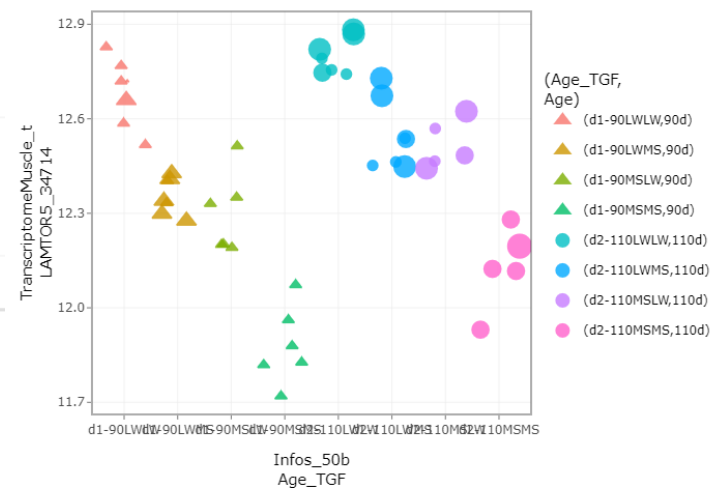
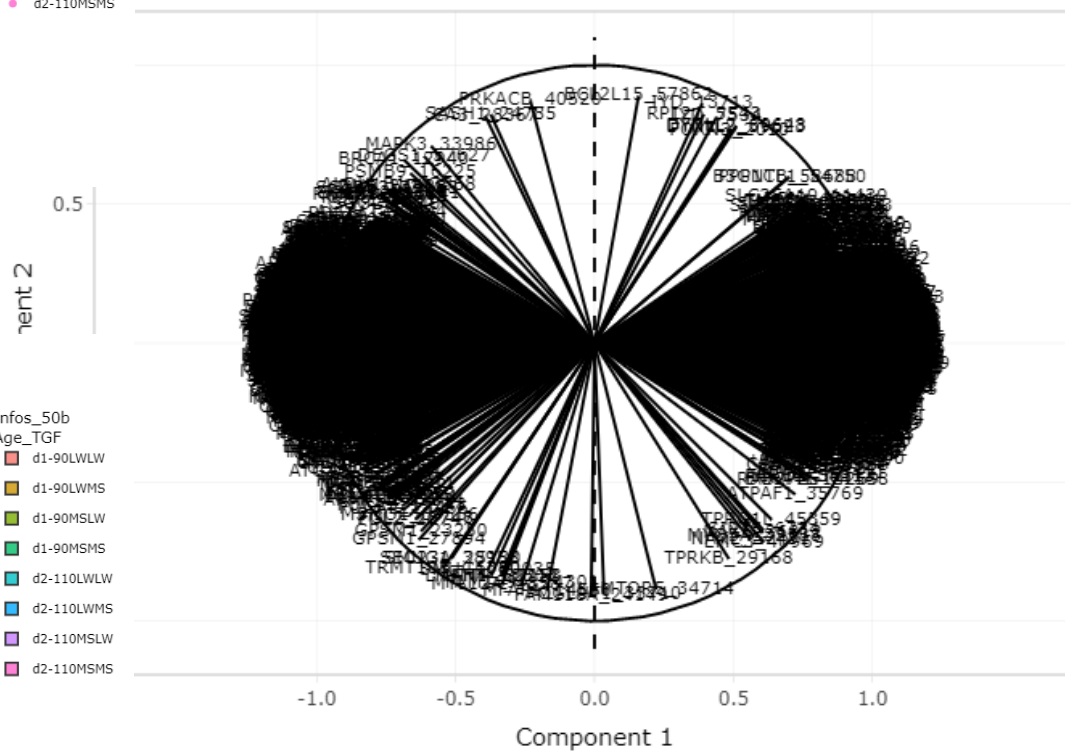
[Preprocessing](#)
[Run PLS-DA](#)
[Explore individuals](#)
[Explore variables](#)
[Extract new data](#)

Age_TGF
8 levels





SDA on TranscriptomeMuscle50_Imputed_2, Infos_50b: correlatio



Differential analysis

[Preprocessing](#)
[Multiple tests](#)
[Posthoc tests](#)
[Extract dataset](#)

1. Run multiple tests ?

Type

parametric
 non-parametric
 automatic

Correction

BH

Threshold

0.05

Run

2. Explore results of multiple tests

[Plots](#)
[Heatmap](#)
[Summary](#)

Test results (Kruskal-Wallis test, corrected with 'BH') for TranscriptomeMuscle_t vs Infos_50b: Age_TGF

| Variables | Test stats. | p-values | Adj. p-values |
|--------------|-------------|--------------|---------------|
| A4GALT_23662 | 36.2485 | 0.000006507 | 0 |
| AAAS_2894 | 34.6425 | 0 | 0 |
| AAAS_18735 | 38.7512 | 0.0000021801 | 0 |
| AAGAB_36620 | 41.0106 | 8.0581e-7 | 0 |
| AARS2_40325 | 35.1548 | 0 | 0 |
| ABCA3_15222 | 36.3064 | 0.0000063452 | 0 |
| ABCA3_58959 | 34.623 | 0 | 0 |

Explore a dataset with PCA

[Run PCA](#)[Explore individuals](#)[Explore variables](#)[Extract new data](#)

1. Display individuals on components ?

Component to display on X (horizontal)

Component to display on Y (vertical)

Color

Infos444_...

Age

Shape

Dataset

Select a variable

Size

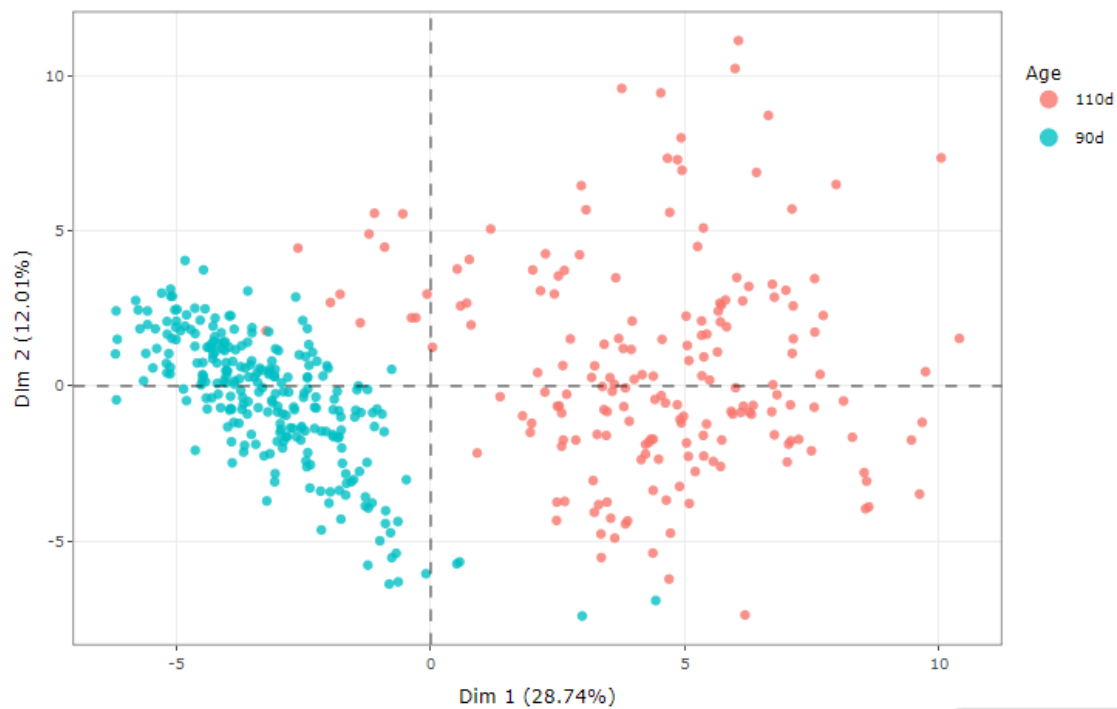
Dataset

Select a variable

Plot individuals

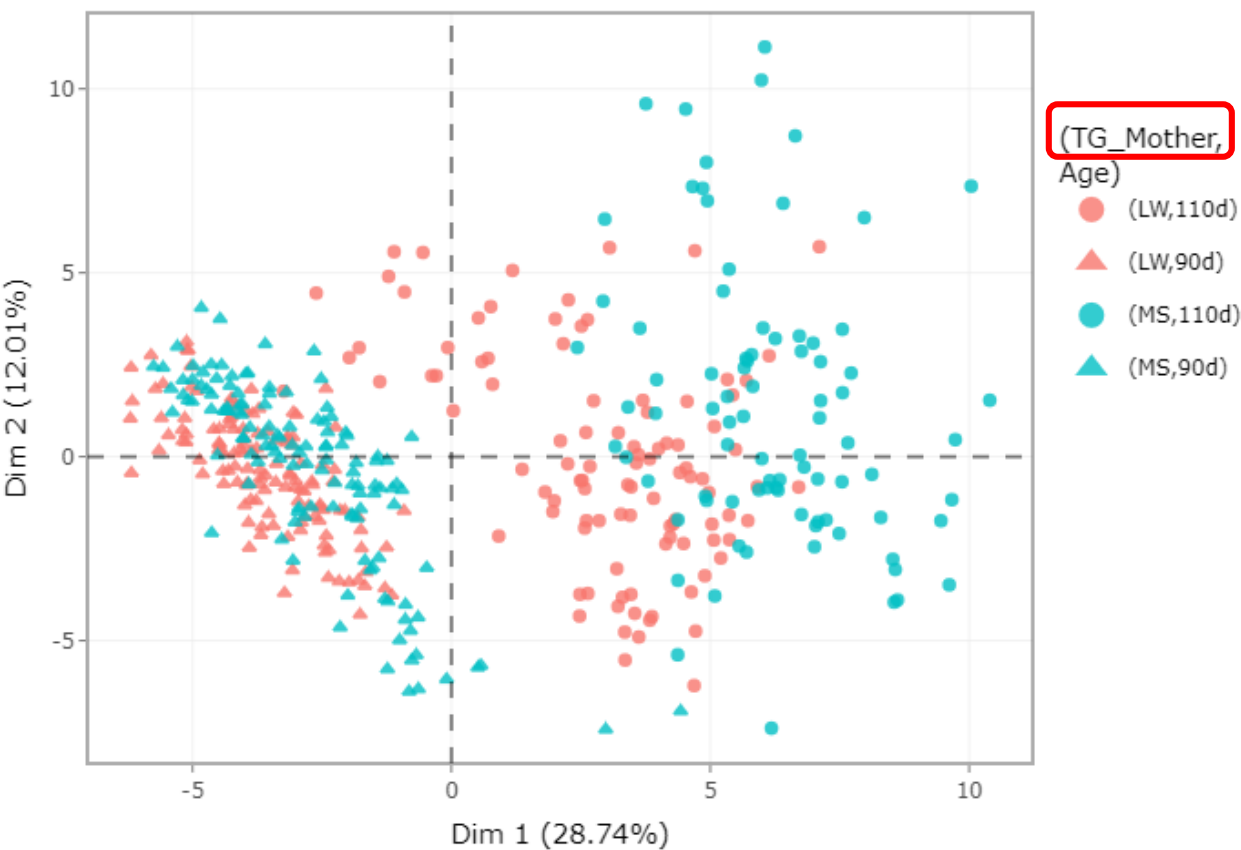
2. Plot individuals of edited_3 on components

PCA on MetabolomePlasma444_t: projection of individuals on the PC

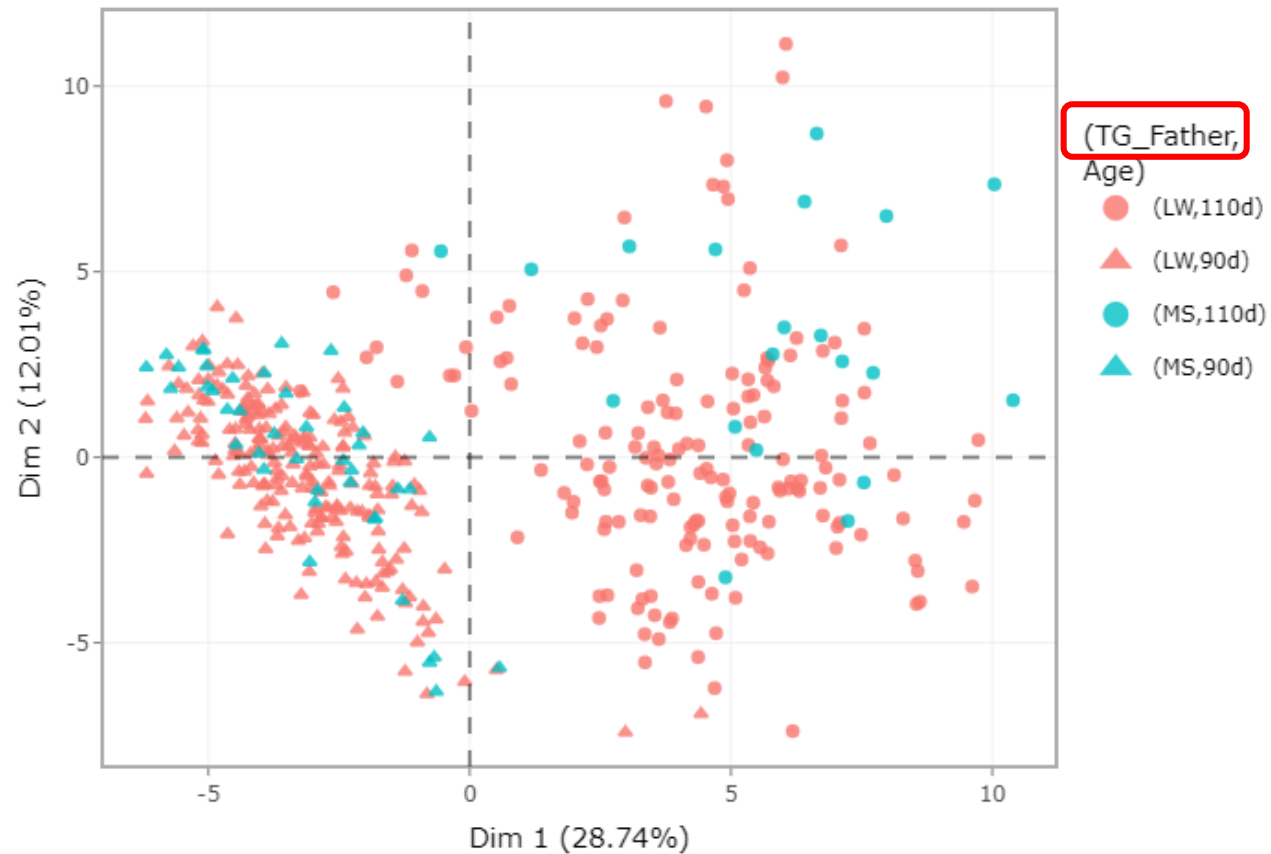


Add to report

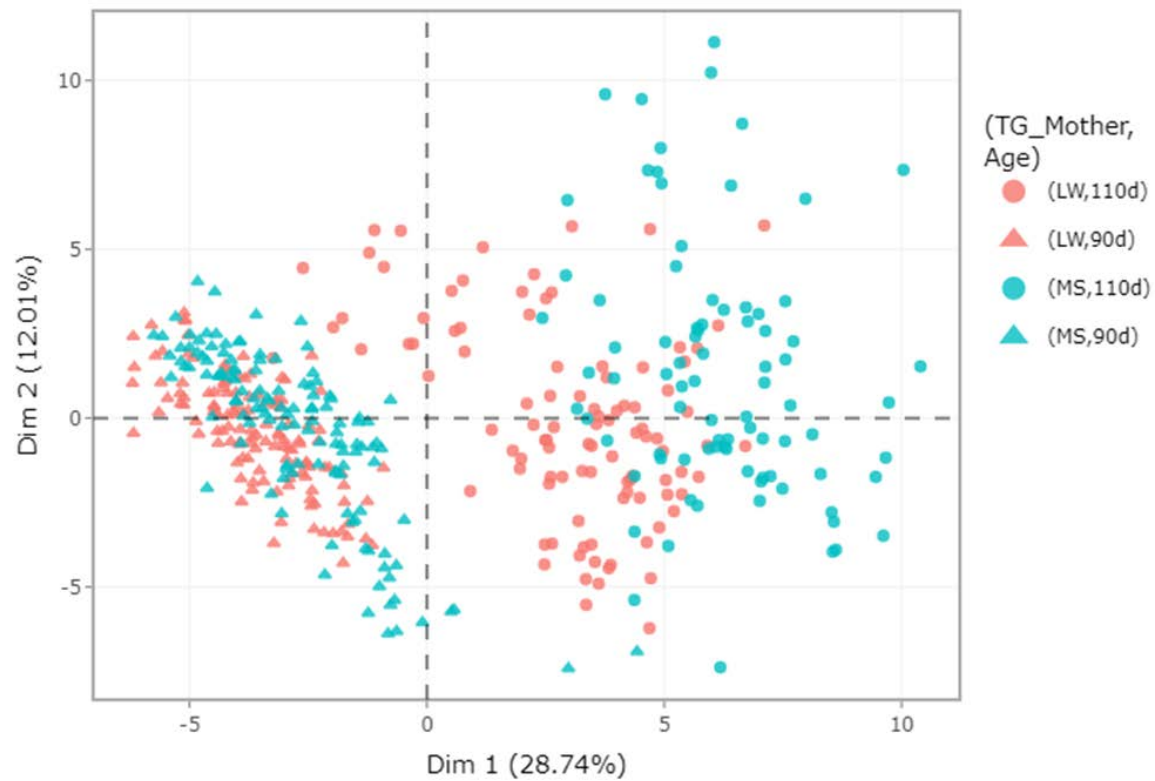
PCA on MetabolomePlasma444_t: projection of individuals on the PC



PCA on MetabolomePlasma444_t: projection of individuals on the PC

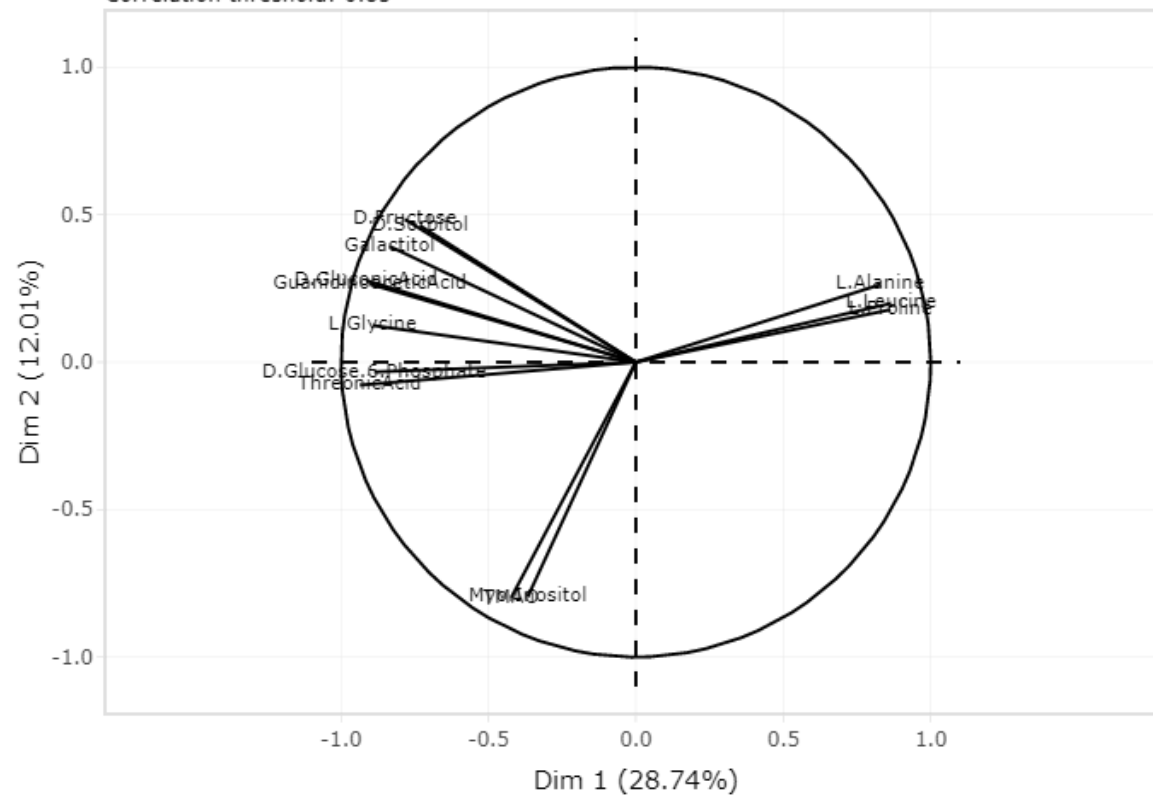


PCA on MetabolomePlasma444_t: projection of individuals on the PC

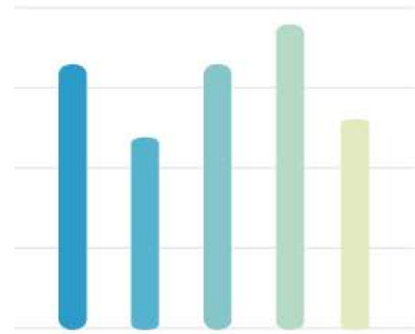


PCA on MetabolomePlasma444_t: correlations of variables with the PCs.

Correlation threshold: 0.85

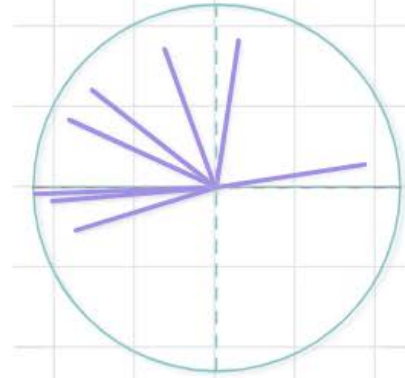


Let's explore a dataset!



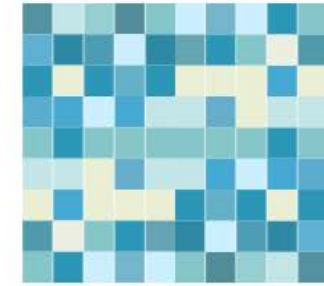
Explore variables in a dataset

Obtain numerical summaries and plots for a few variables.



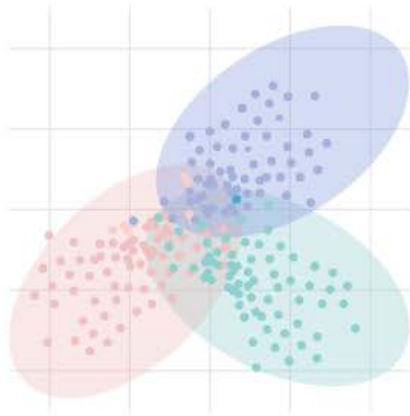
Explore a dataset with PCA

Perform Principal Component Analysis on a dataset.



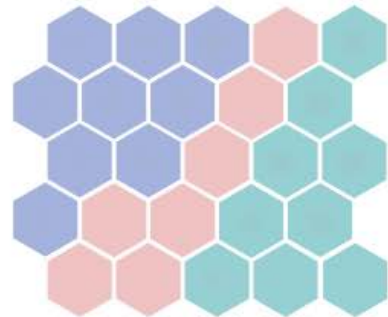
Explore a dataset with a heatmap

Obtain the heatmap of a dataset.



Clustering

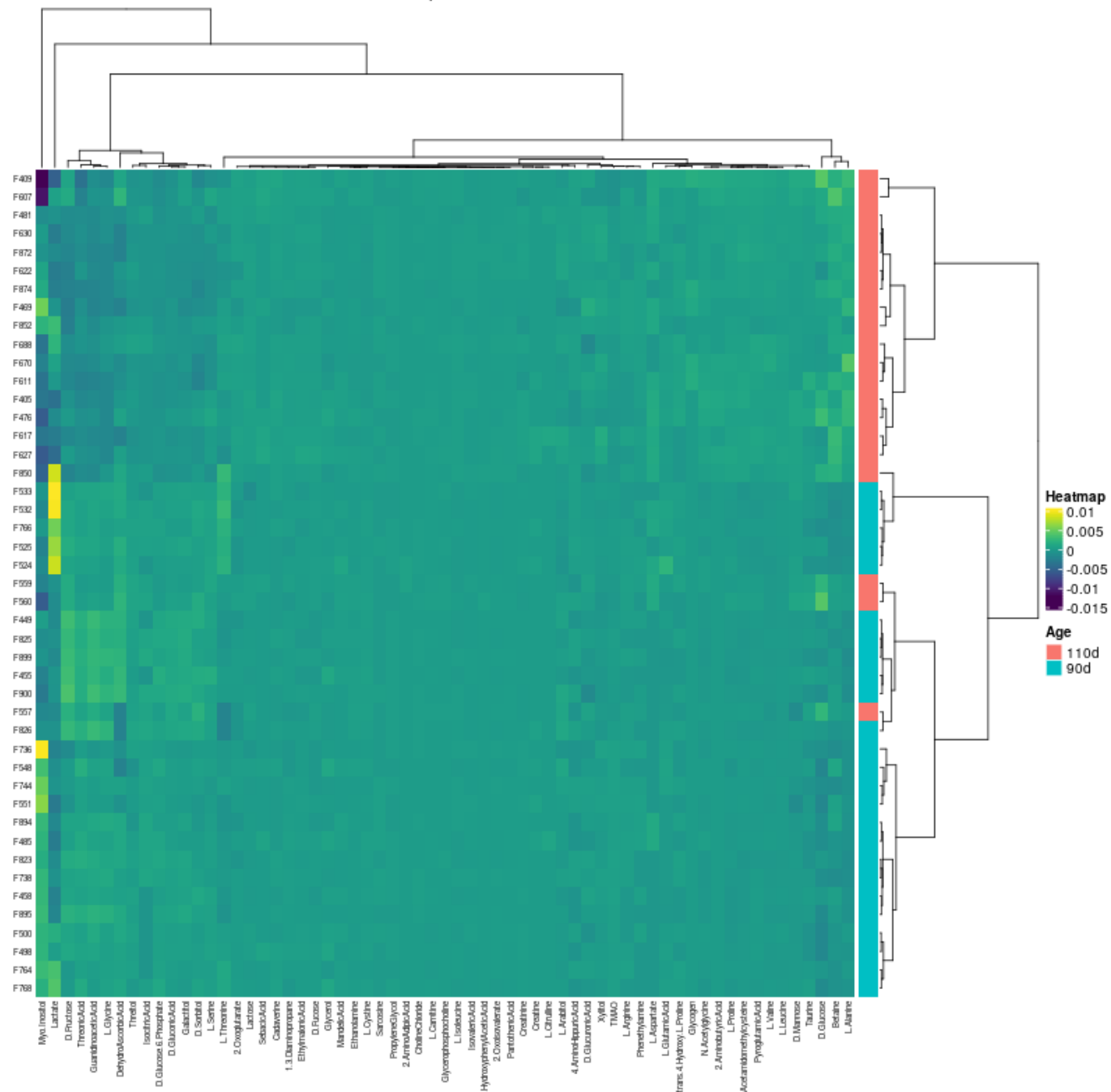
Cluster the individuals of a dataset.



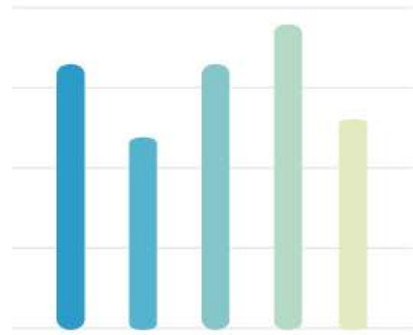
Self-Organizing Map

Use SOM as a clustering and visualization method.

Heatmap of dataset

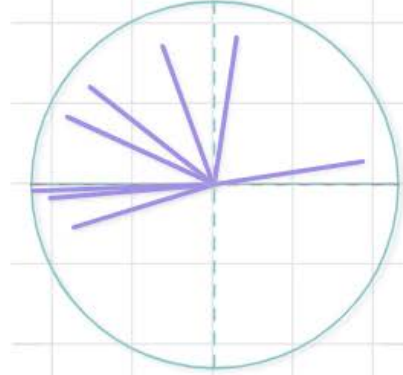


Let's explore a dataset!



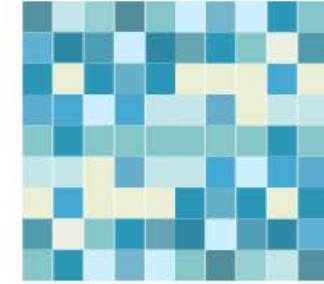
Explore variables in a dataset

Obtain numerical summaries and plots for a few variables.



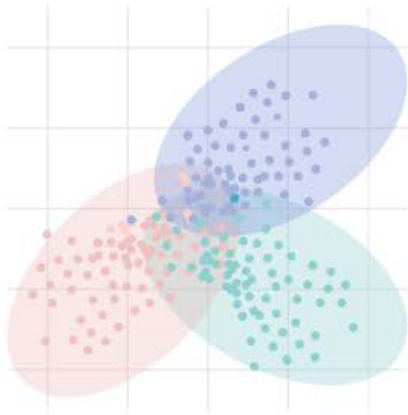
Explore a dataset with PCA

Perform Principal Component Analysis on a dataset.



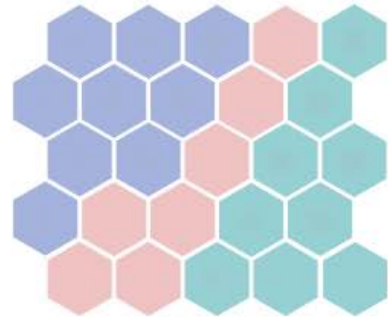
Explore a dataset with a heatmap

Obtain the heatmap of a dataset.



Clustering

Cluster the individuals of a dataset.



Self-Organizing Map

Use SOM as a clustering and visualization method.

Self-Organizing Map

[Self-Organizing Map](#)
[Explore individuals](#)
[Explore prototypes](#)
[Superclustering](#)

1. Run a SOM analysis ?

Select a dataset

Proteome_Muscle_50 ▾

Topology

hexagonal ▾

Map length

5

Map width

5

Seed

21171

You can fix the random seed used to ensure reproducibility of the analysis.

[Run SOM](#)

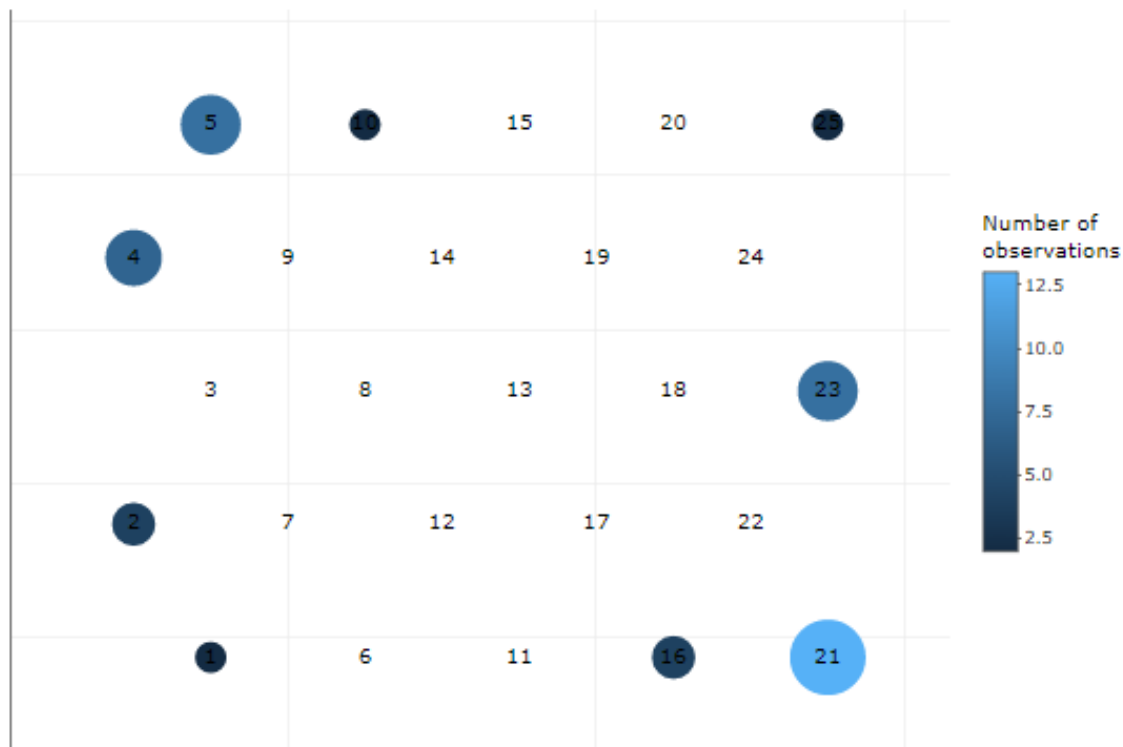
2. Explore SOM clustering

[Plots](#)

[Summary](#)

[Cluster view](#)

Proteome_Muscle_50: distribution of individuals 📷 📊



Self-Organizing Map

[Self-Organizing Map](#)
[Explore individuals](#)
[Explore prototypes](#)
[Superclustering](#)

Quality criteria of the SOM map

| Topographic error | Quantization error |
|-------------------|--------------------|
| 0.04 | 781.6191 |

Correlation ratio

| individual | Ratio |
|----------------------|--------|
| <input type="text"/> | |
| P0457 | 0.9942 |
| P0390 | 0.9939 |
| ACO2_0718 | 0.9933 |
| P1008 | 0.9932 |
| P1057 | 0.9921 |
| P0240 | 0.9919 |
| TMM3 Isoform 7_1058 | 0.9914 |

[Self-organizing map](#)
[Summary](#)
[Cluster view](#)


Quality criteria:

- *topographic error*: is a measure of how well the map is organized (*i.e.*, if two clusters are close, can you trust the fact that corresponding individuals are similar?). This number is always between 0 (best possible organization) and 1 (worst possible organization), with 0 being the value expected for small maps;
- *quantization error*: is a measure of the clustering quality (very similar to within-dispersion in clustering). This number is always non negative (the smaller its value, the better the clustering).



Correlation ratio: This table displays the 20 first variables with the largest percentage of inertia reproduced by the clusters of the map. The largest the percentage of inertia, the more relevant the corresponding variable is to explain differences between clusters.

Self-Organizing Map

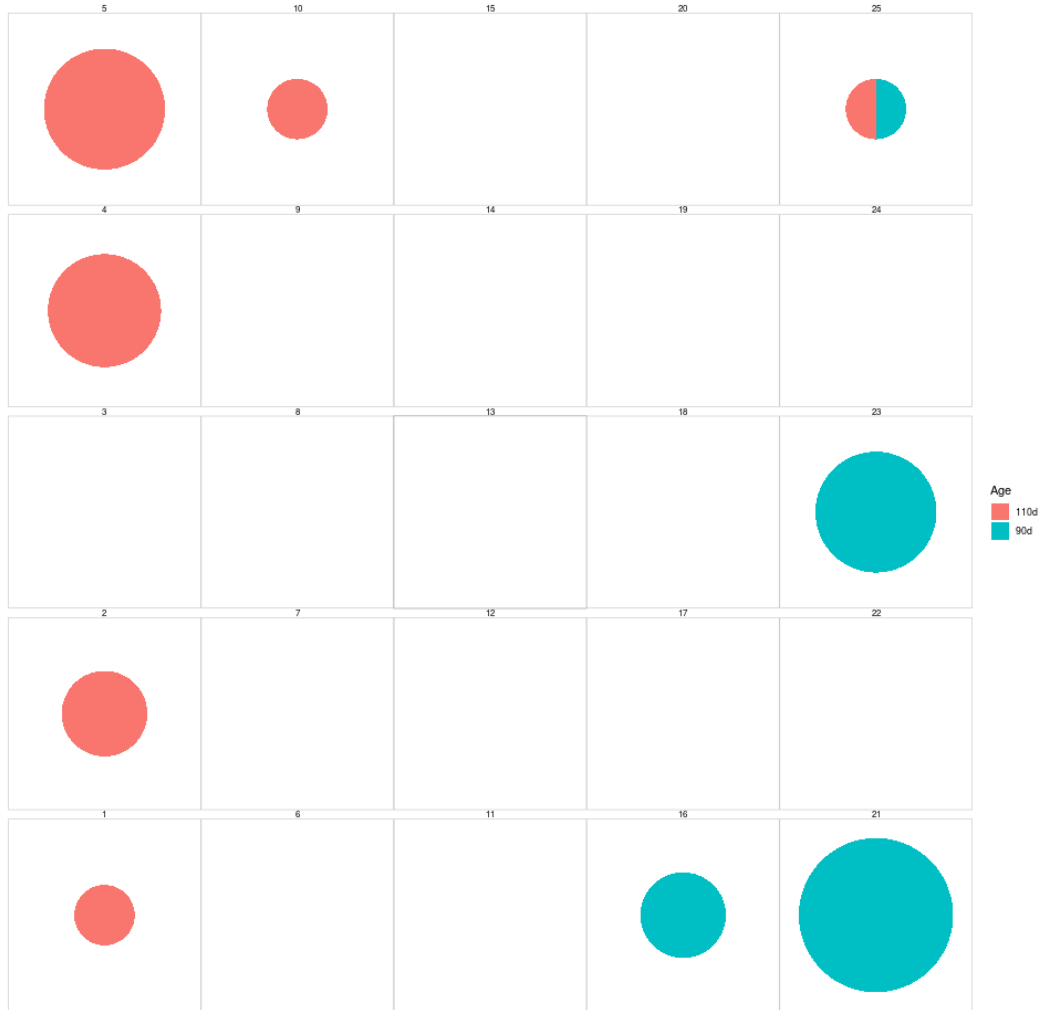
Self-Organizing Map

Explore individuals

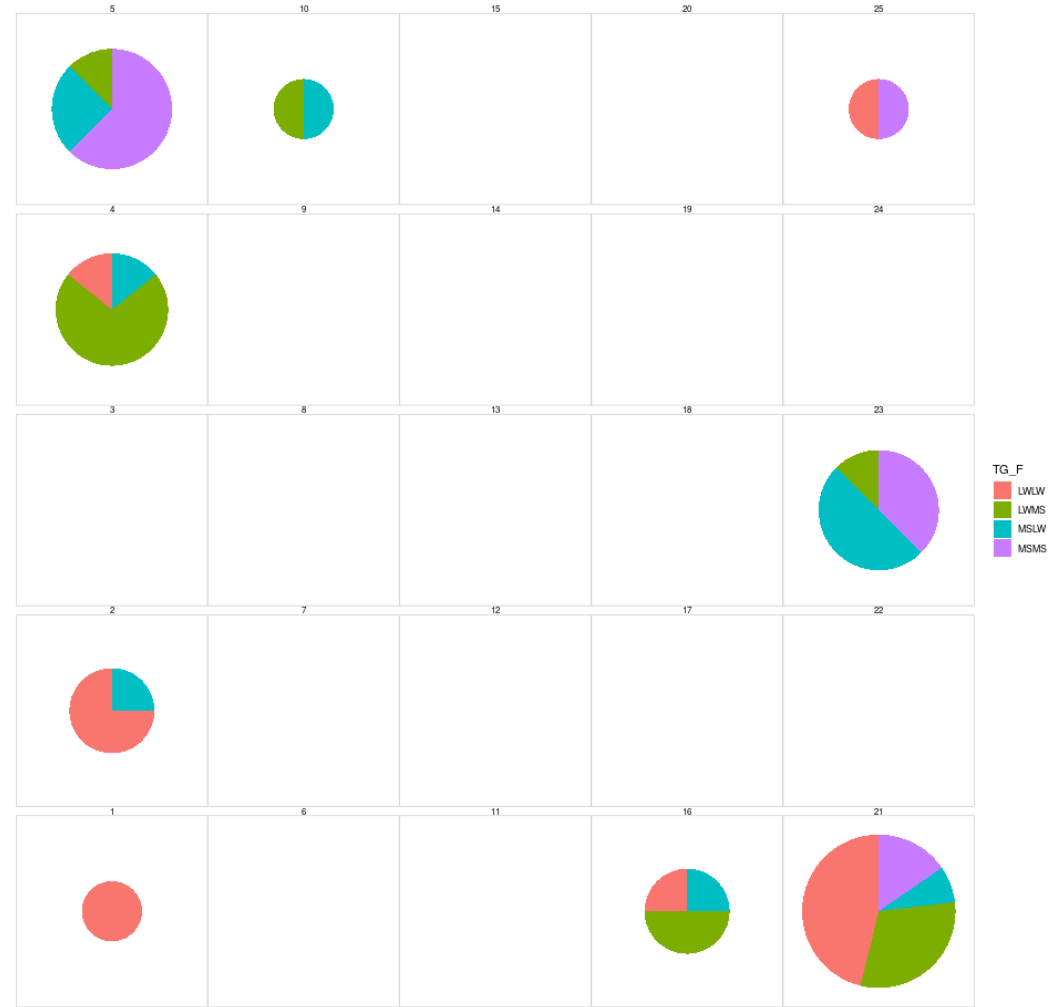
Explore prototypes

Superclustering

SOM on Proteome_Muscle_50: additional variable overview



SOM on Proteome_Muscle_50: additional variable overview



Self-Organizing Map

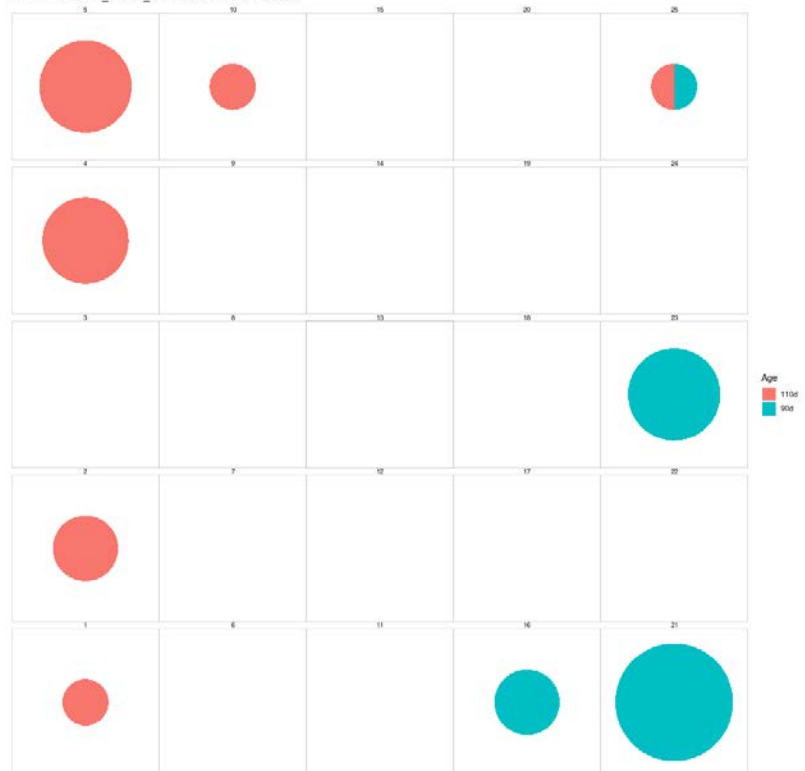
Self-Organizing Map

Explore individuals

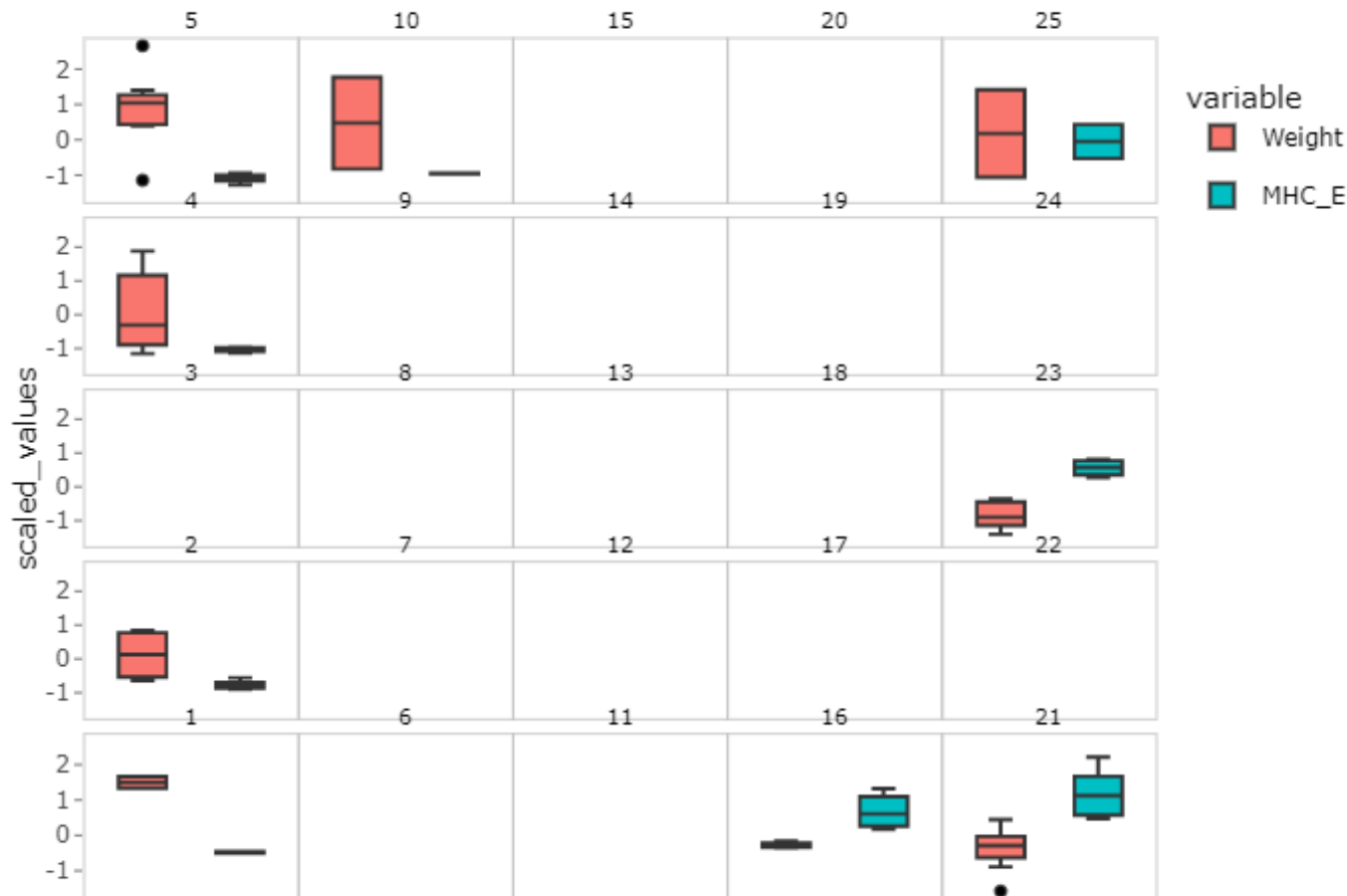
Explore prototypes

Superclustering

SOM on Proteome_Muscle_50: additional variable overview



SOM on Proteome_Muscle_50: additional variable overview



Self-Organizing Map

[Self-Organizing Map](#)
[Explore individuals](#)
[Explore prototypes](#)
[Superclustering](#)

1. Display prototypes



Plot type

Color



Select one variable

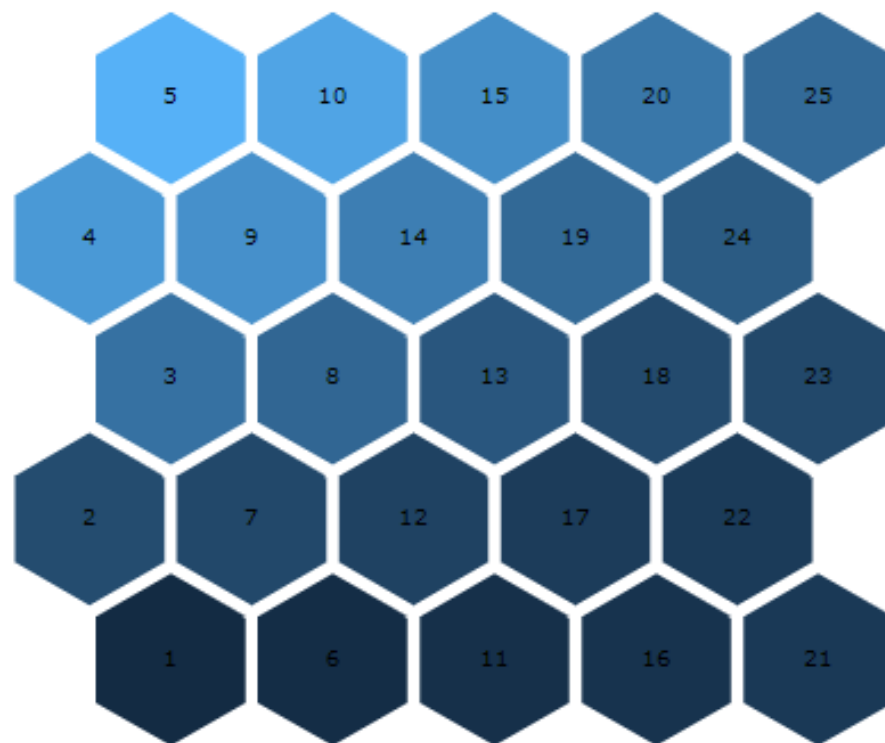
GPD1..Isoform.4._1032



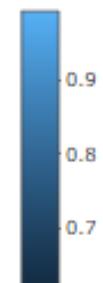
Explore individuals

2. Explore prototypes

SOM on Proteome_Muscle_50: prototypes overview



value of
GPD1..Isoform.4._1032
for each prototype



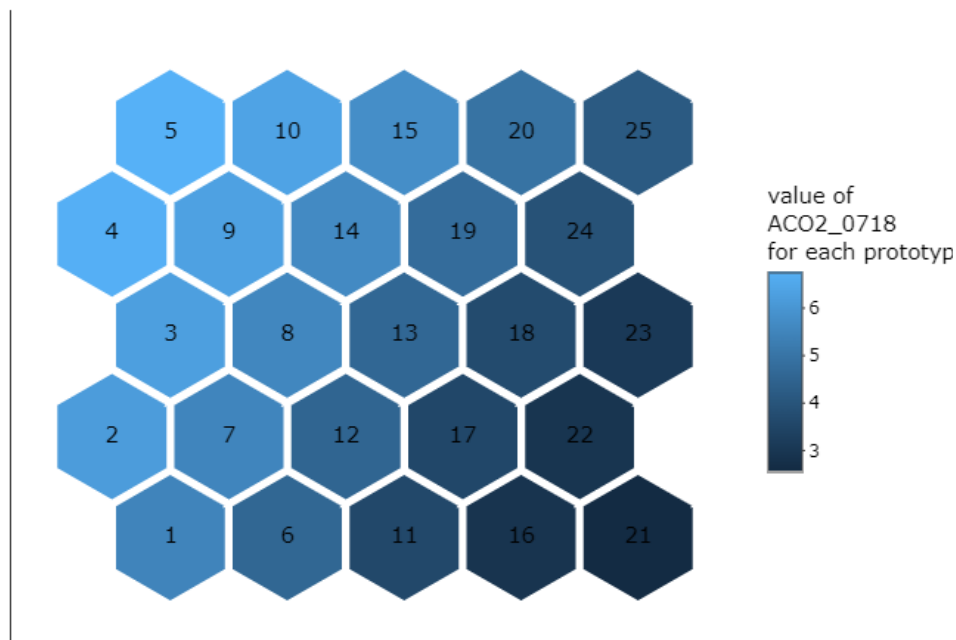
Add to report

Self-Organizing Map

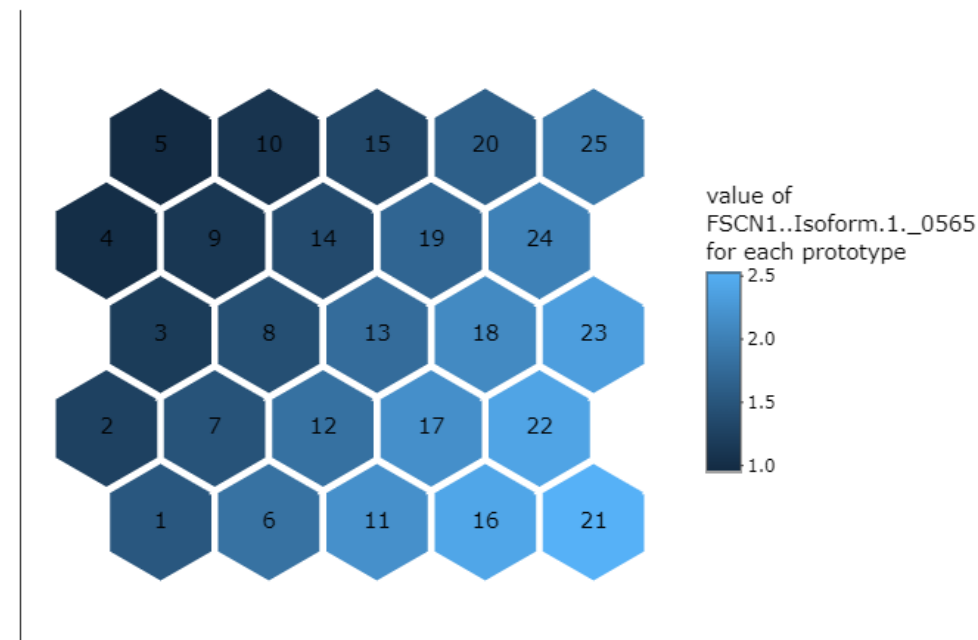
[Self-Organizing Map](#)
[Explore individuals](#)
[Explore prototypes](#)
[Superclustering](#)

| | Corr.Ratio |
|------------------------|------------|
| P0457 | 0.9942 |
| P0390 | 0.9939 |
| ACO2_0718 | 0.9933 |
| P1008 | 0.9932 |
| P1057 | 0.9921 |
| P0240 | 0.9919 |
| TNNT3..Isoform.7._1058 | 0.9914 |
| FH_0451 | 0.9911 |
| P0913 | 0.9911 |
| P0773 | 0.991 |
| CKMT2_0429 | 0.9908 |
| P0719 | 0.9907 |
| NDUFV1_0496 | 0.9906 |
| ATP5A1_0520 | 0.9904 |
| P0269 | 0.9892 |
| PGK1..Isoform.1._0427 | 0.9891 |
| MDH2_0275 | 0.988 |
| FSCN1..Isoform.1._0565 | 0.9875 |
| ACADVL_1051 | 0.9867 |
| P0276 | 0.9867 |

SOM on Proteome_Muscle_50: prototypes overview



SOM on Proteome_Muscle_50: prototypes overview

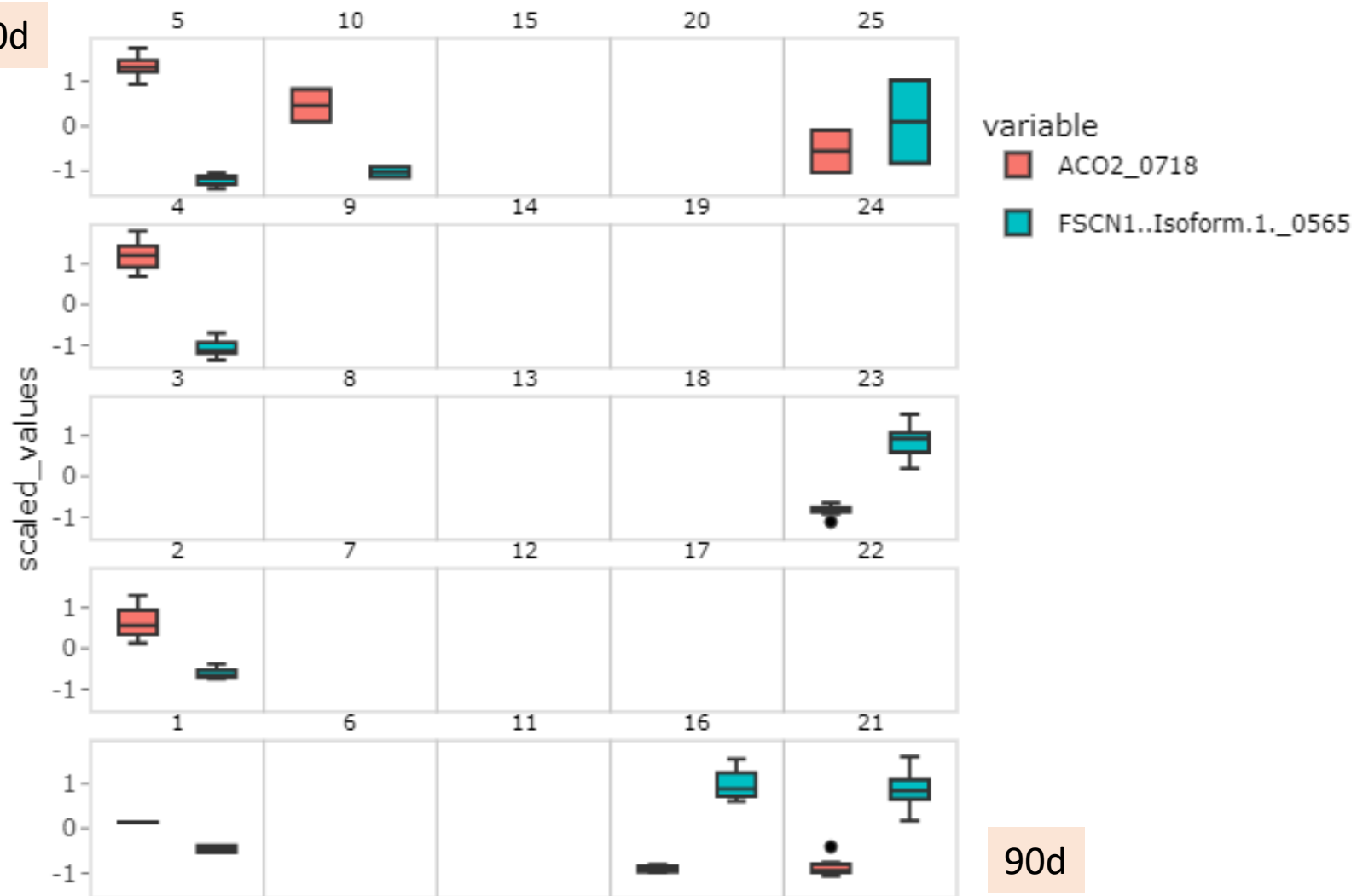


Self-Organizing Map

[Self-Organizing Map](#)
[Explore individuals](#)
[Explore prototypes](#)
[Superclustering](#)

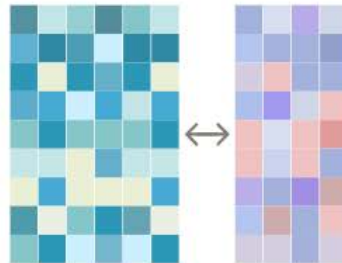
SOM on Proteome_Muscle_50: overview of individuals

110d



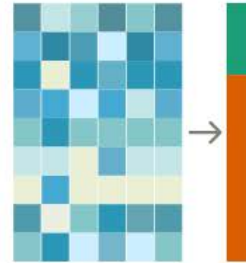
| | Corr.Ratio |
|------------------------|------------|
| P0457 | 0.9942 |
| P0390 | 0.9939 |
| ACO2_0718 | 0.9933 |
| P1008 | 0.9932 |
| P1057 | 0.9921 |
| P0240 | 0.9919 |
| TNNT3..Isoform.7._1058 | 0.9914 |
| FH_0451 | 0.9911 |
| P0913 | 0.9911 |
| P0773 | 0.991 |
| CKMT2_0429 | 0.9908 |
| P0719 | 0.9907 |
| NDUFV1_0496 | 0.9906 |
| ATP5A1_0520 | 0.9904 |
| P0269 | 0.9892 |
| PGK1..Isoform.1._0427 | 0.9891 |
| MDH2_0275 | 0.988 |
| FSCN1..Isoform.1._0565 | 0.9875 |
| ACADVL_1051 | 0.9867 |
| P0276 | 0.9867 |

Let's integrate data!



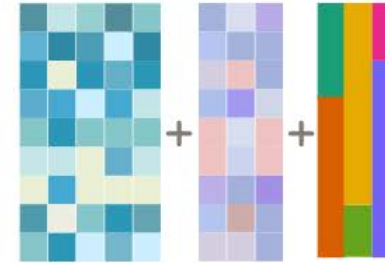
Integrate two datasets with PLS

Performs Partial Least Squares analysis on two datasets.



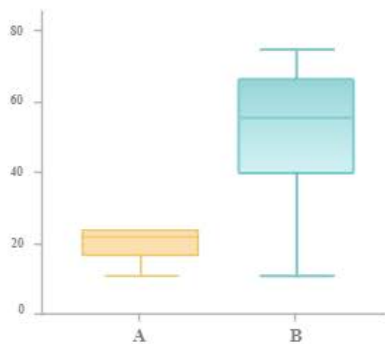
Integrate two datasets with PLS-DA

Performs Partial Least Squares analysis on two datasets.



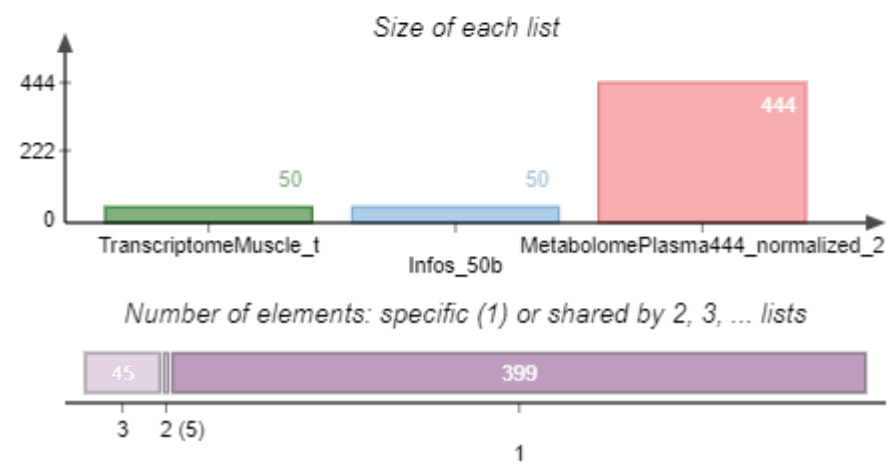
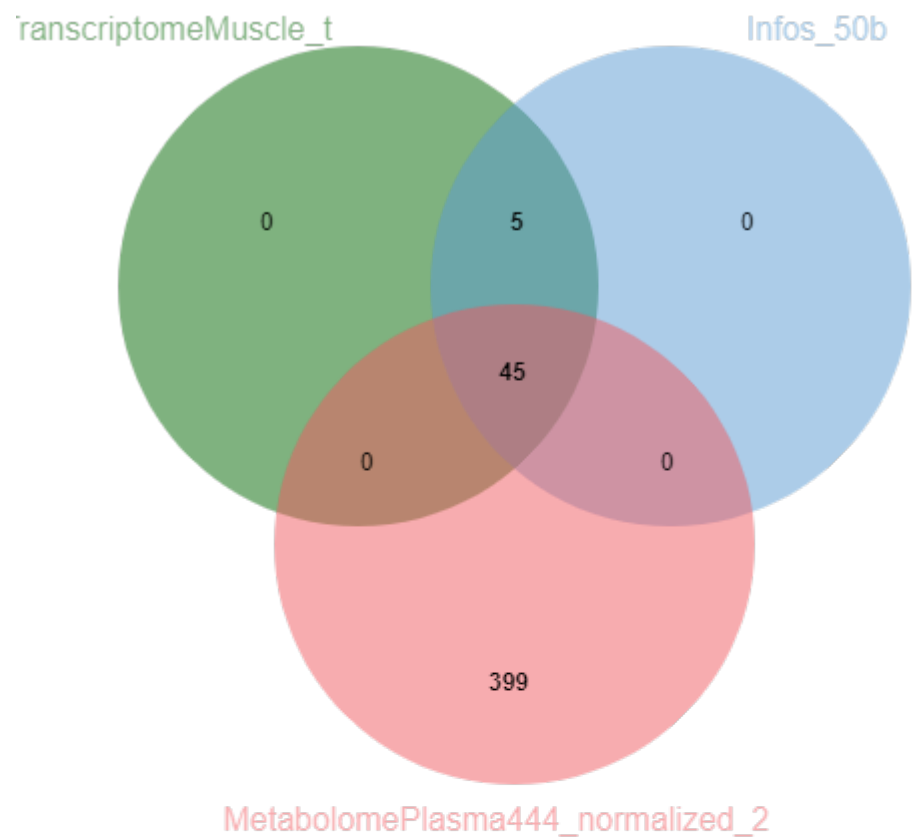
Integrate datasets with MFA

Perform Multiple Factor Analysis on several datasets.



Differential analysis

Perform differential analysis for all numeric variables of a dataset.



Integrate datasets with MFA

Preprocessing

Run MFA

Explore individuals

Explore variables

Explore groups

Extract new data

1. Display individuals on components



Component to display on X (horizontal)

1 ✓

Component to display on Y (vertical)

2 ✓

Color

Info_50b_... TG_F

Shape

Info_50b_... Age

Size

Info_50b_... Weight

Partial individual group

Partial individual group

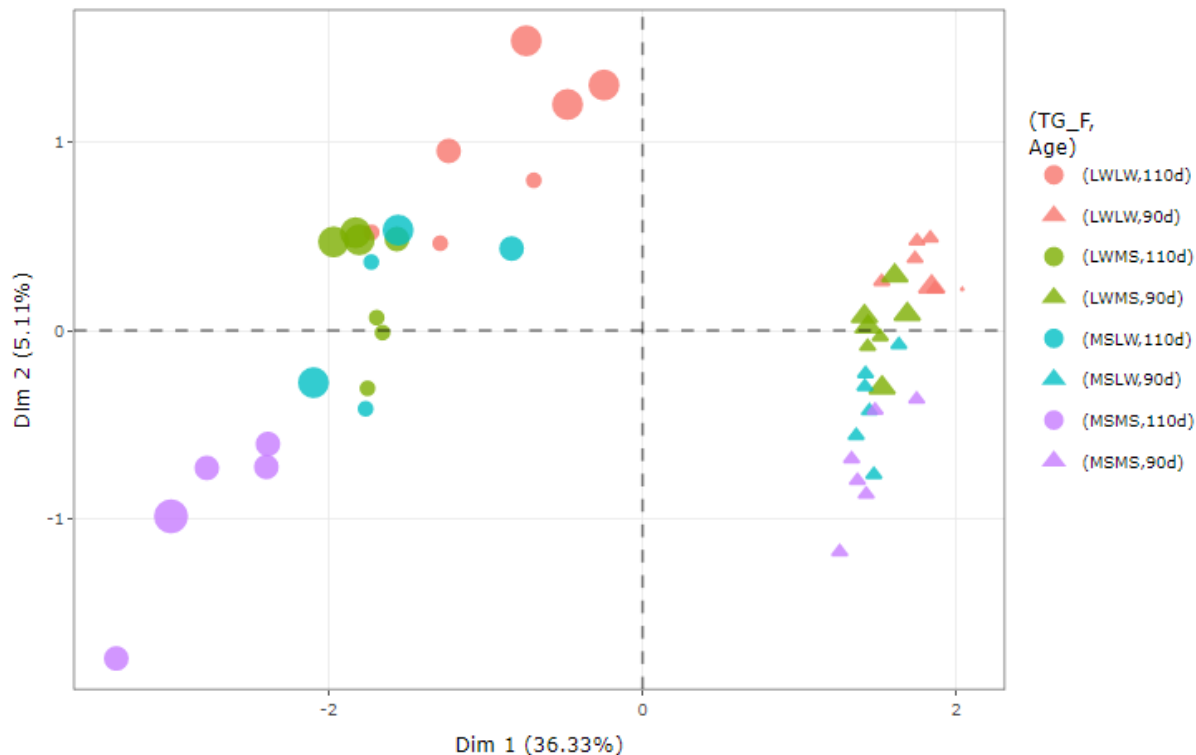
Partial individuals

▶ Add an individual

Plot individuals

2. Plot individuals on components

MFA on Info_50b_e, TranscriptomeMuscle50_Imputed_1, Proteome_Muscle_50: projection o



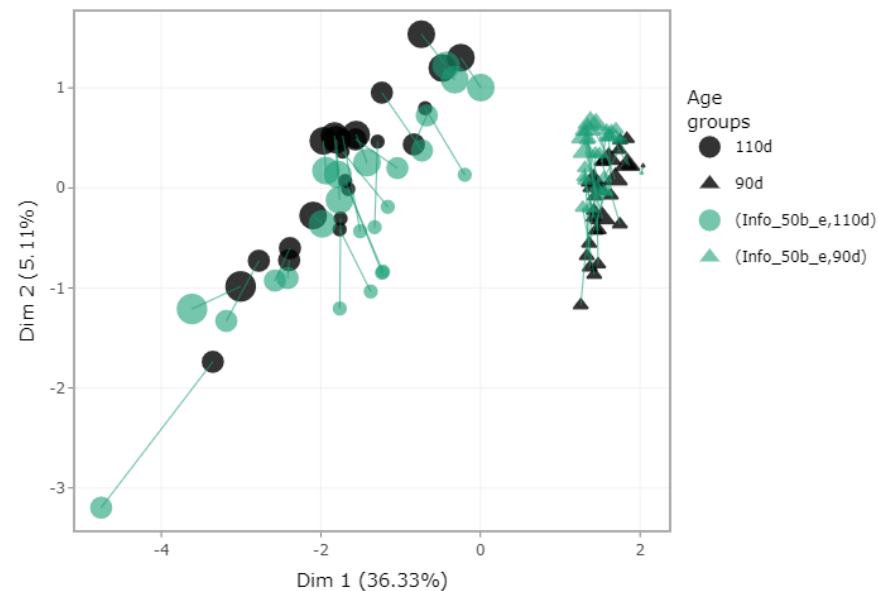
Add to report

Integrate datasets with MFA

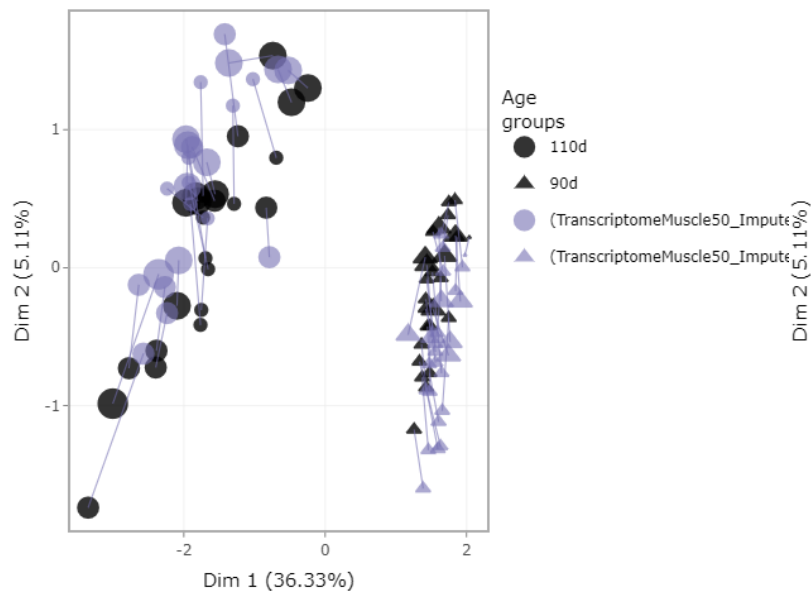
[Preprocessing](#)
[Run MFA](#)
[Explore individuals](#)
[Explore variables](#)
[Explore groups](#)
[Extract new data](#)

Next steps...

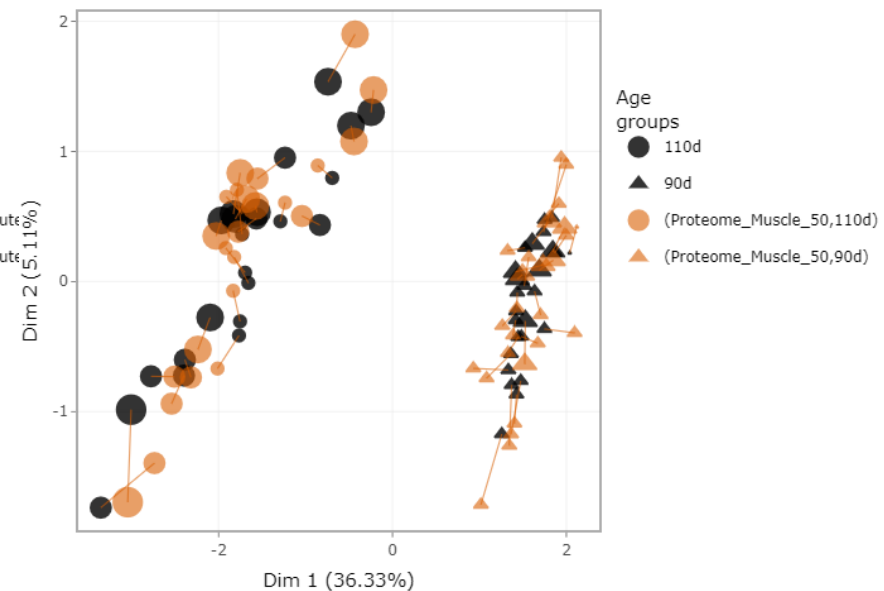
MFA on Info_50b_e, TranscriptomeMuscle50_Imputed_1, Proteome_Muscl



MFA on Info_50b_e, TranscriptomeMuscle50_Imputed_1, Proteo



MFA on Info_50b_e, TranscriptomeMuscle50_Imputed_1, Proteome_Muscl



Datasets 8

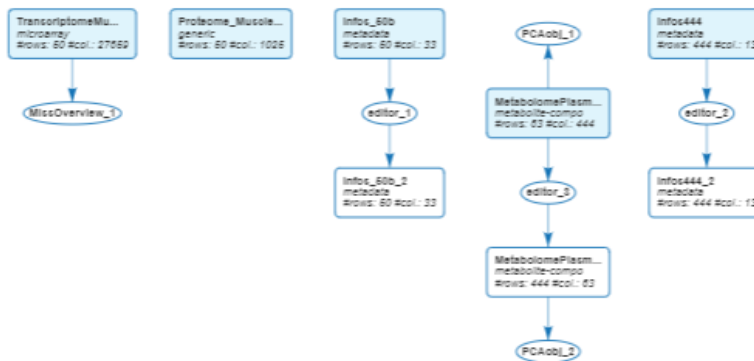
Variables 29,283

Analyses 6

Remaining days

Copy url

Workflow



WinZip (Non enregistré) - asterics_reports1d94db1c2ee.zip

Fichier Actions Options Aide

Nouveau Ouvrir Favoris Ajouter Extraire Voir Contrôle Assistant

| Nom | Type | Modifié | Taille | Taux | Compressé |
|---|----------------------|------------------|-----------|------|-----------|
| astericsreport editor_11d91a2b5246.html | Chrome HTML Document | 17/08/2022 09:13 | 1 964 748 | 73% | 528 764 |
| astericsreport editor_21d95b0edb02.html | Chrome HTML Document | 17/08/2022 09:13 | 1 964 745 | 73% | 528 769 |
| astericsreport editor_31d94bd4355e.html | Chrome HTML Document | 17/08/2022 09:13 | 1 964 636 | 73% | 528 729 |
| astericsreport MissOverview_11d923bfd802.html | Chrome HTML Document | 17/08/2022 09:13 | 6 232 916 | 71% | 1 819 891 |
| astericsreport PCAobj_11d97665a528.html | Chrome HTML Document | 17/08/2022 09:13 | 5 418 126 | 71% | 1 547 641 |
| astericsreport PCAobj_21d95ea9658d.html | Chrome HTML Document | 17/08/2022 09:13 | 5 522 510 | 72% | 1 568 596 |

Sélectionnés 0 fichiers, 0 octets Total 6 fichiers, 22 528Ko

All analyses

Remove Export report ?

| <input checked="" type="checkbox"/> | Name | Function |
|---------------------------------------|----------------|--------------------|
| > <input checked="" type="checkbox"/> | editor_1 | r_edit_dataset |
| > <input checked="" type="checkbox"/> | editor_2 | r_edit_dataset |
| > <input checked="" type="checkbox"/> | MissOverview_1 | r_missing_overview |
| > <input checked="" type="checkbox"/> | PCAobj_1 | r_famd |
| > <input checked="" type="checkbox"/> | editor_3 | r_edit_dataset |
| > <input checked="" type="checkbox"/> | PCAobj_2 | r_famd |

Merci à toute l'équipe d'Asterics
Merci pour votre attention



To be continued...